



# USING eDNA ANALYSIS TO DETERMINE THE PRESENCE OF AQUATIC SPECIES

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5<sup>th</sup> Field Technologies for Data Collection in Forestry, Fisheries,  
and Natural Resources  
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# Outline

- **Overview**
- **Discerning salmon redds using eDNA**
- **eDNA as an index of fish abundance**
- **Protocols**
- **Resources**

# Aquatic survey methods



# Aquatic survey methods



Electrofishing



Snorkeling

# Aquatic survey methods



Electrofishing



Snorkeling



eDNA













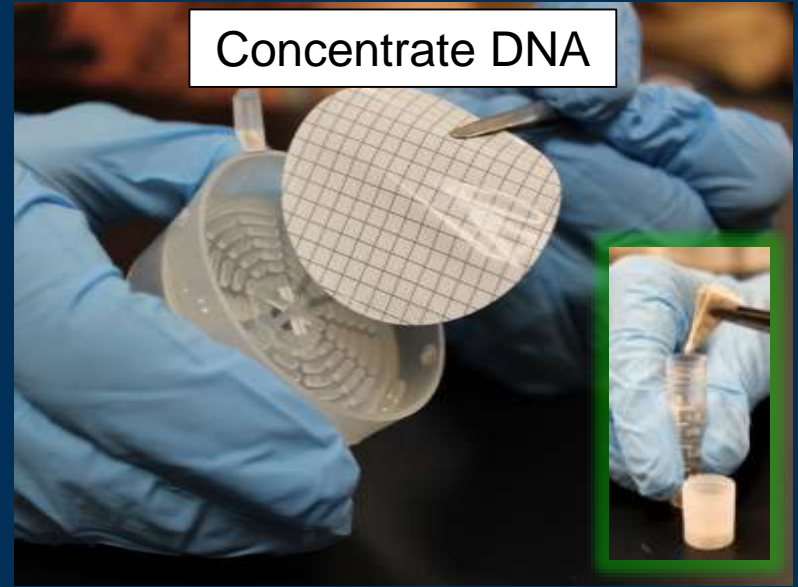


# The Basic Approach

Collect Water Sample



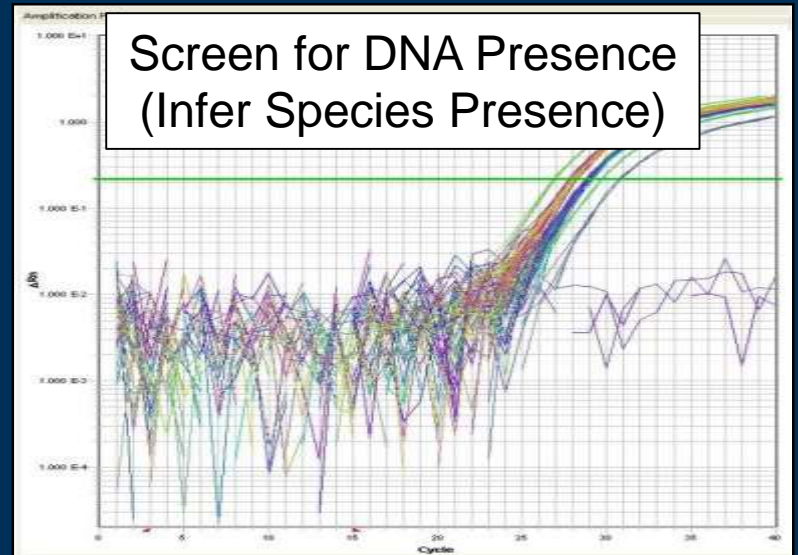
Concentrate DNA



Extract DNA (all) and Amplify Target Sequence

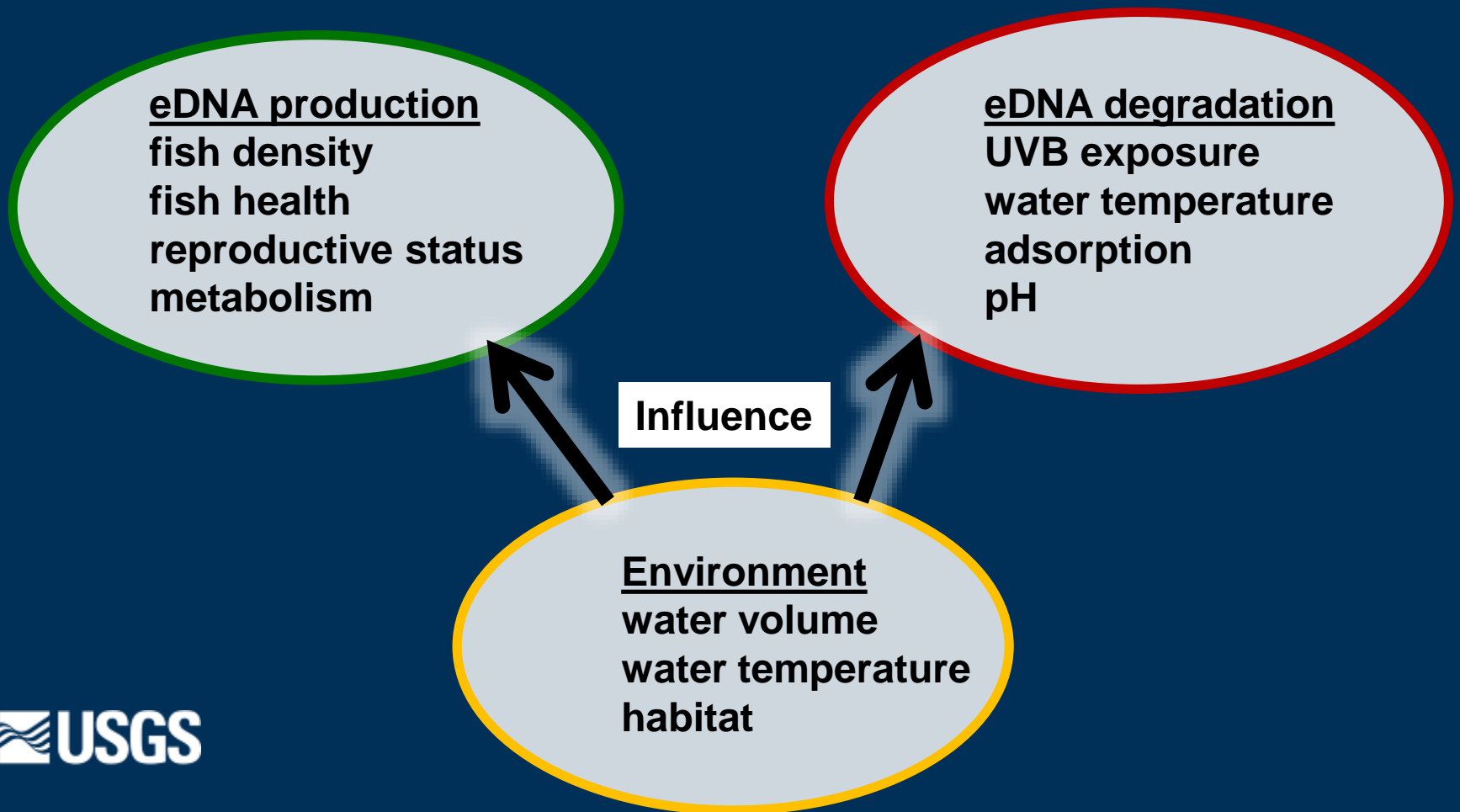


Screen for DNA Presence (Infer Species Presence)

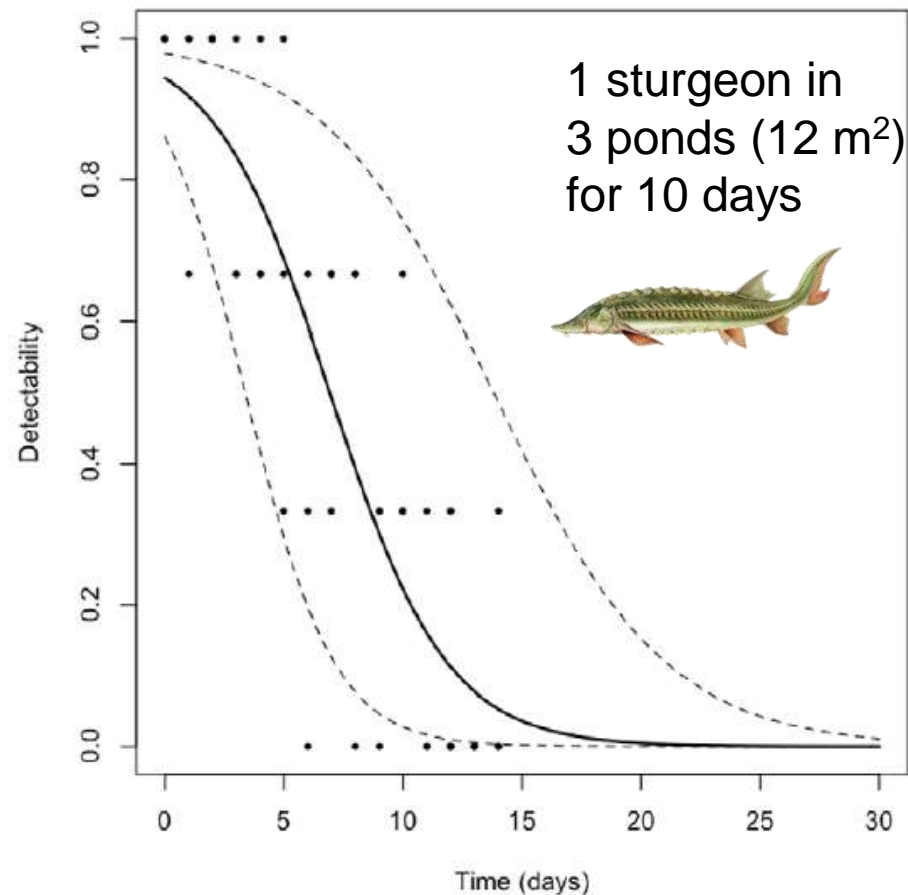
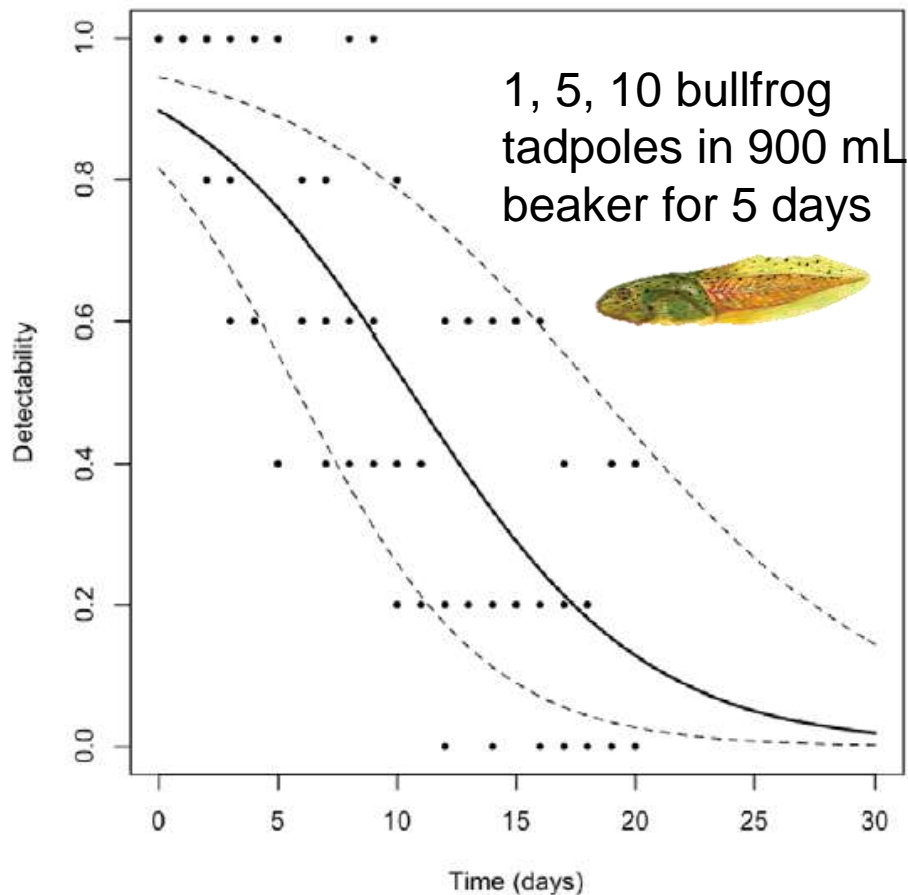


# [eDNA] = production - degradation

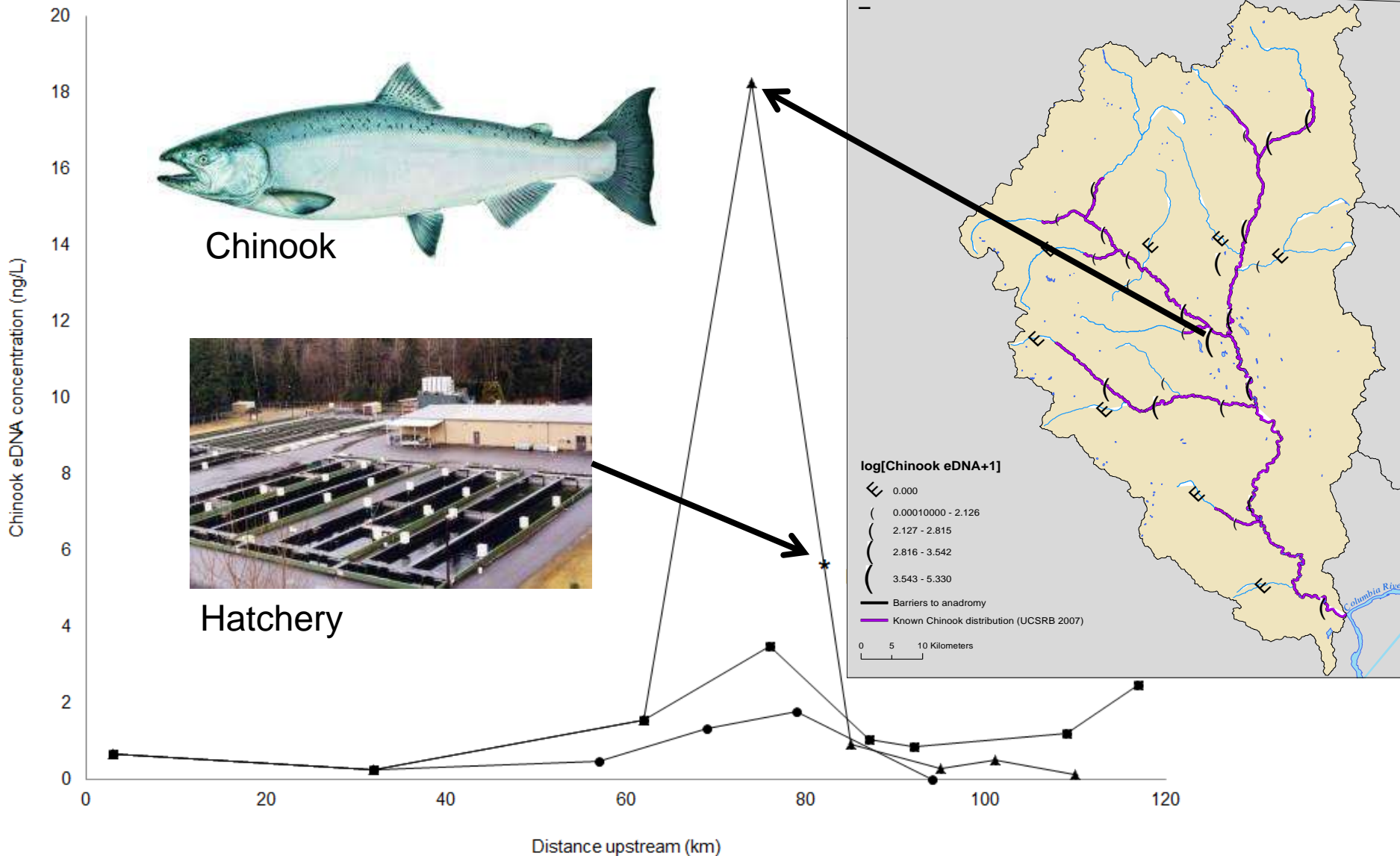
Example: fish



# How long does DNA persist in water?



# Where should samples be collected?



# Integrating into Existing Monitoring Programs

## Traditional survey methods

<u>Species density</u>		<u>Effort required for detection</u>
High	←————→	Low
Low	←————→	High

## eDNA survey methods

<u>Species density</u>		<u>Effort required for detection</u>
High	←————→	Low
Low	←————→	Low

\*Highly sensitive eDNA methods could be useful alternative to investing high effort



# Integrating into Existing Monitoring Programs

## Traditional survey methods



## eDNA survey methods



\*Highly sensitive eDNA methods could be useful alternative to investing high effort

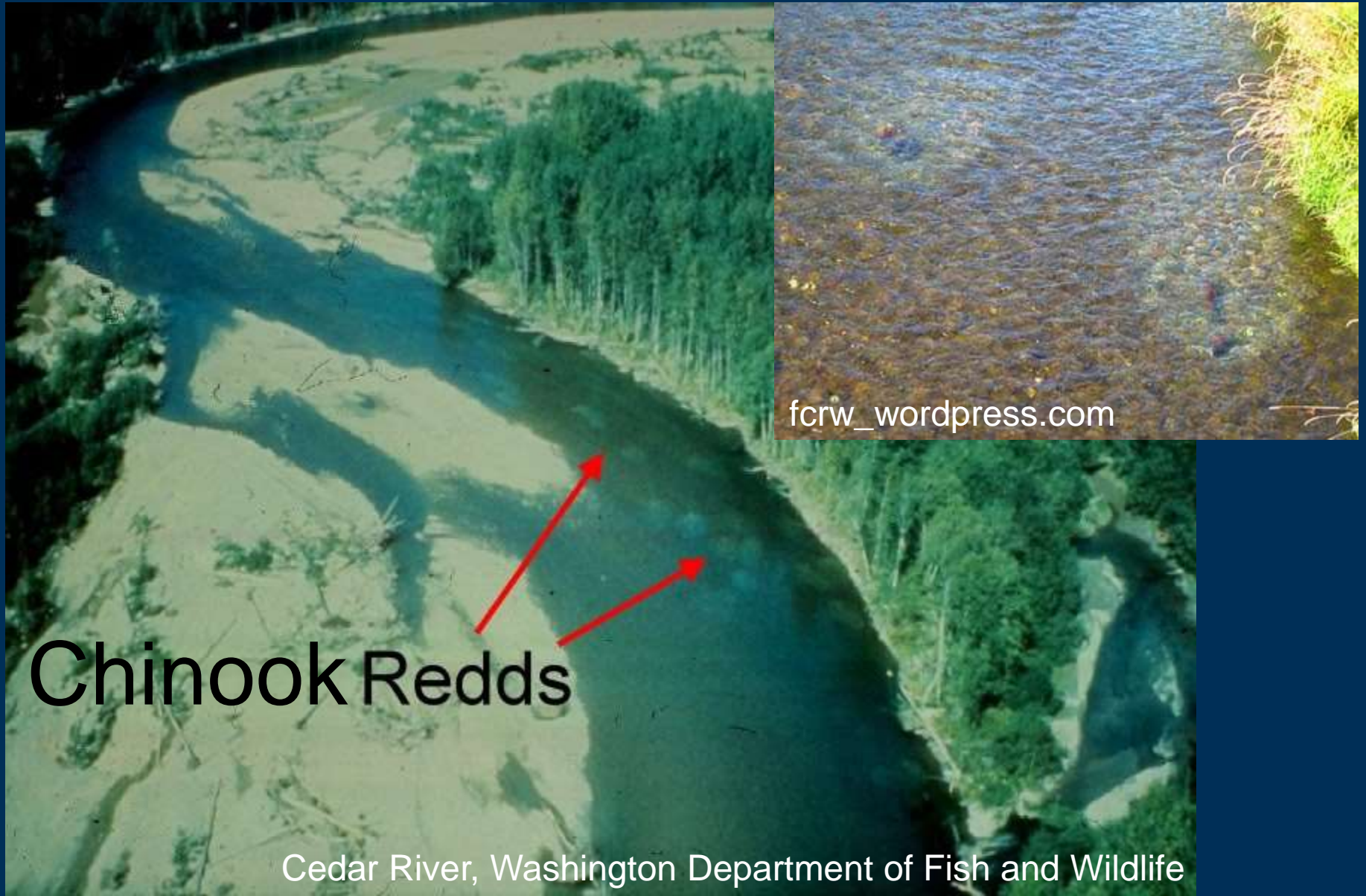


# Monitoring Salmon Populations



Photo used with permission; © Brian Miller (CCT/OBMEP)

# Monitoring Salmon Populations



Chinook Redds

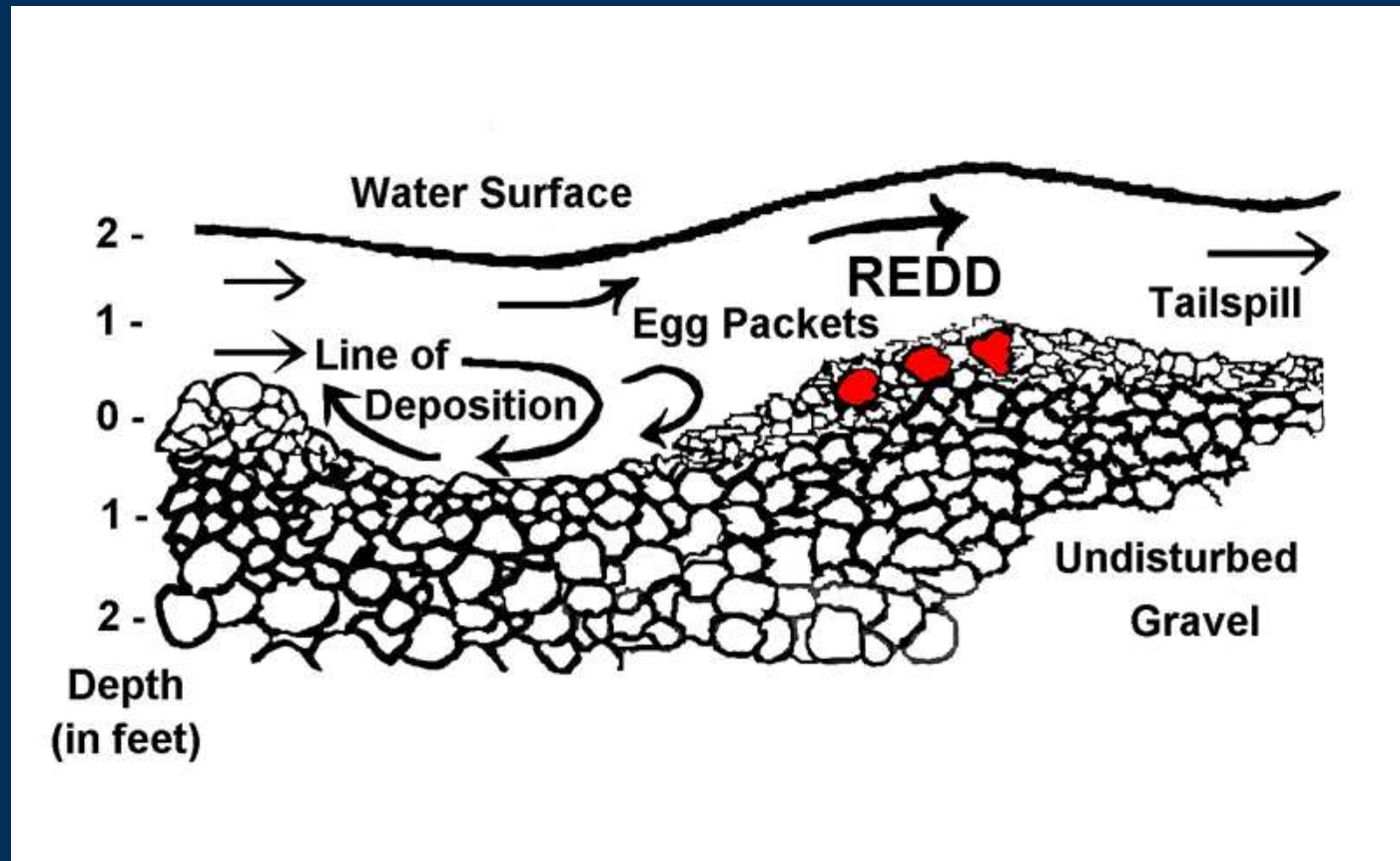
Cedar River, Washington Department of Fish and Wildlife

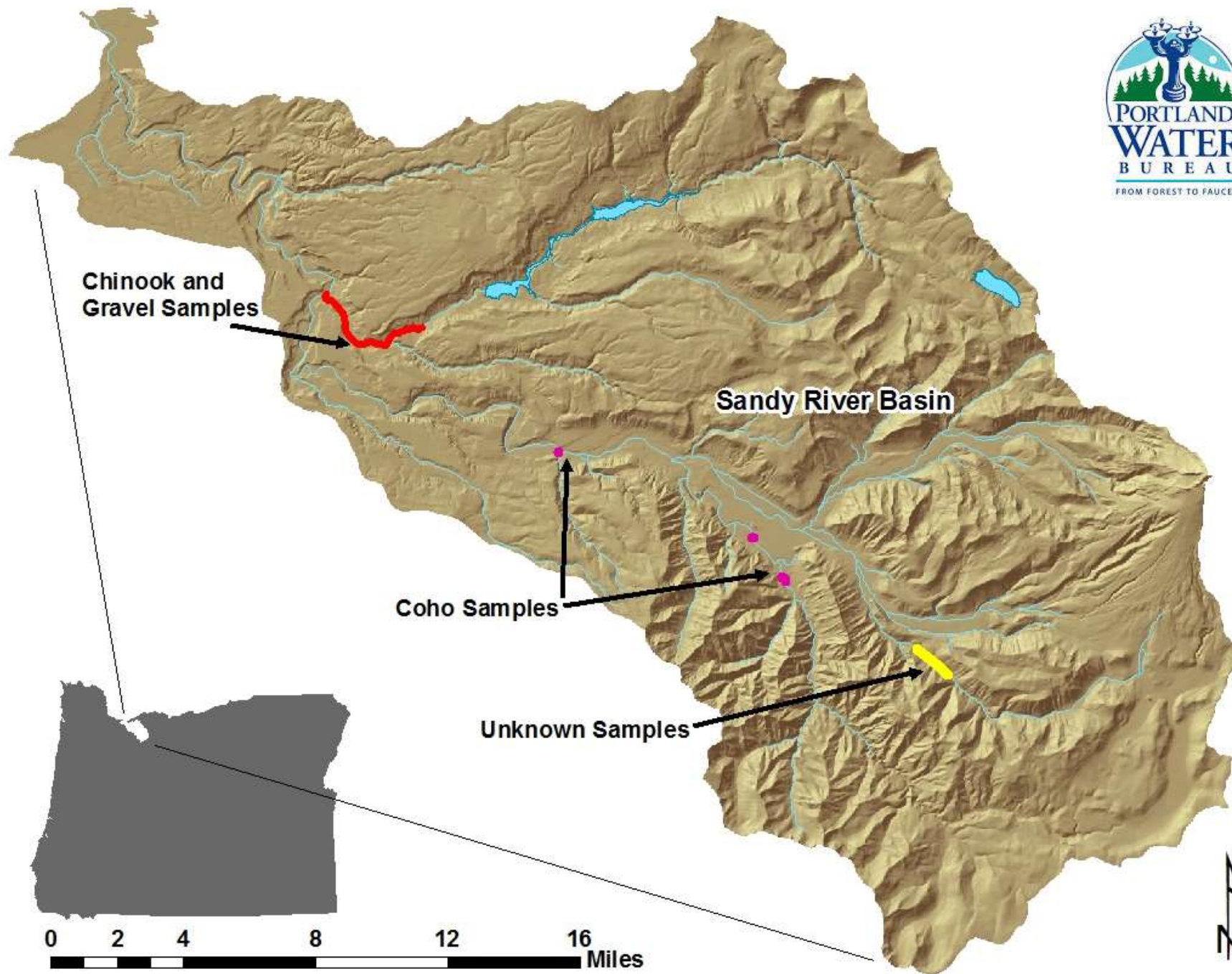
# Habitat & timing used to differentiate redds where species co-occur



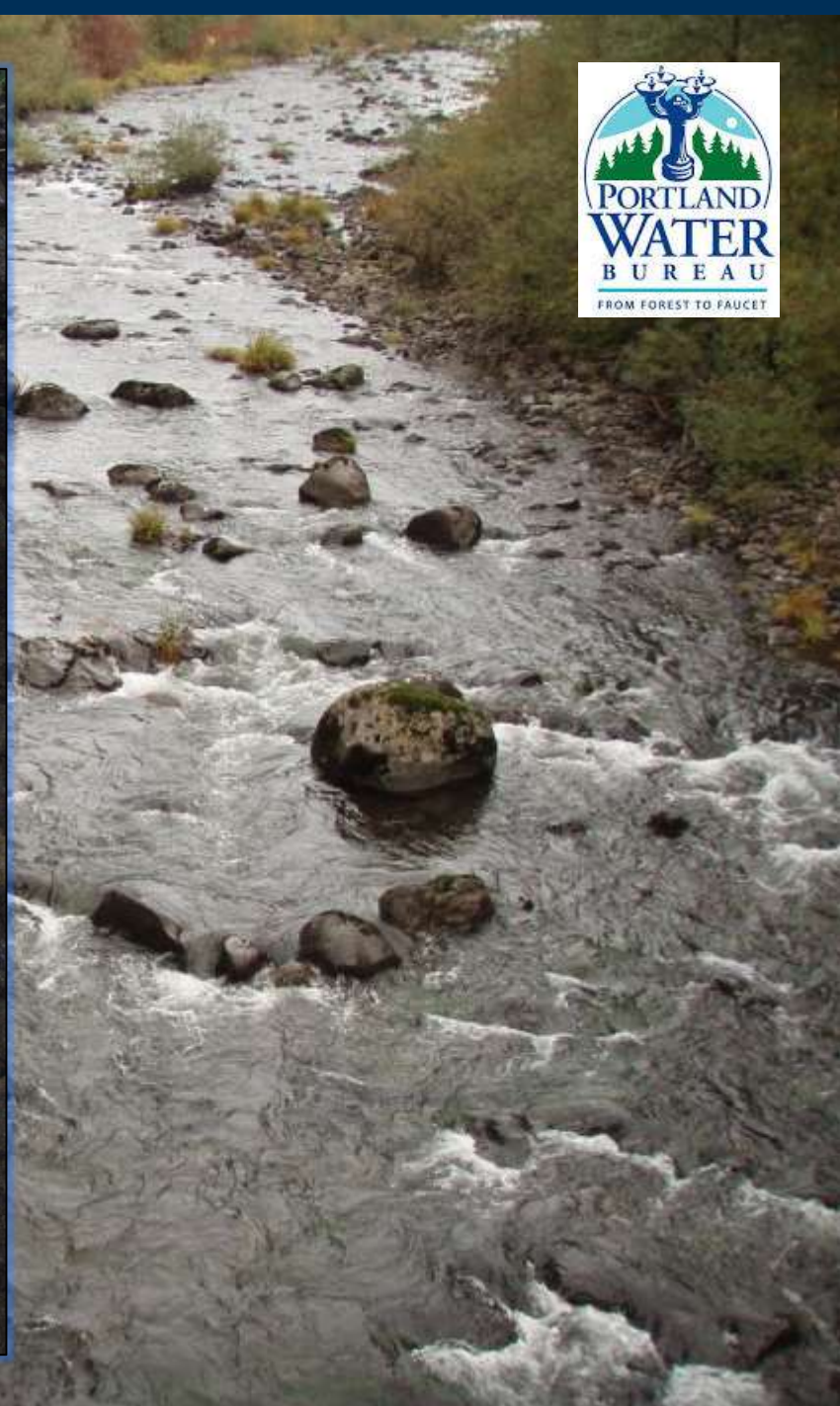


1. How much salmon DNA is in the environment (water column and gravel) during spawning?
2. Can we differentiate coho redds from chinook redds using eDNA analysis?



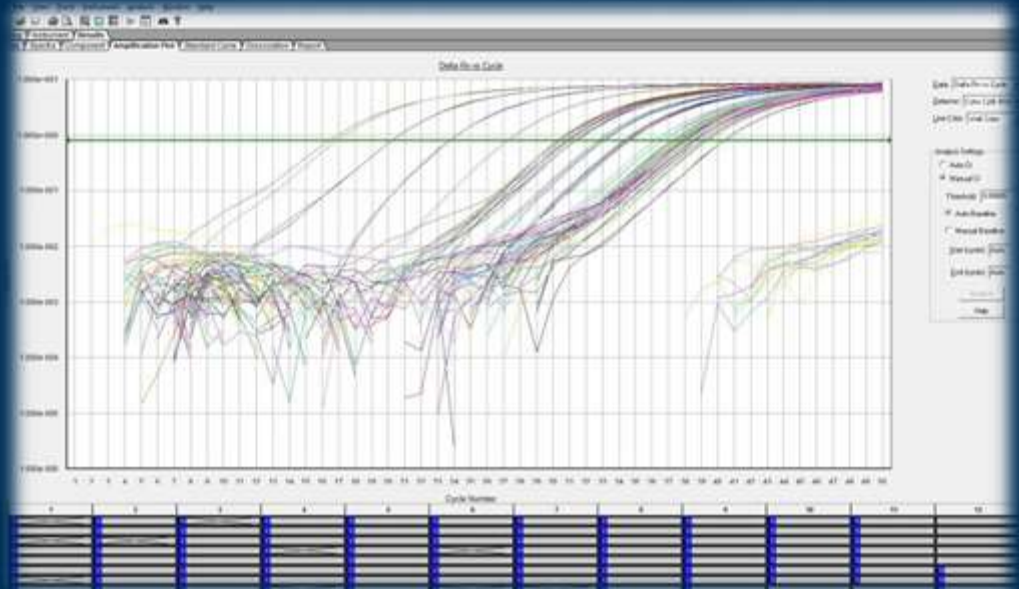


**Burke Strobel,  
Portland Water Bureau**

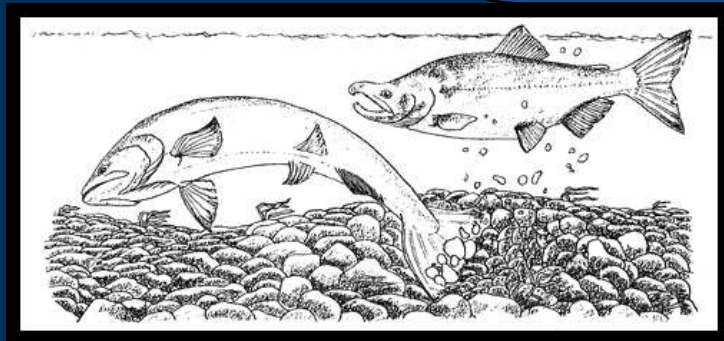
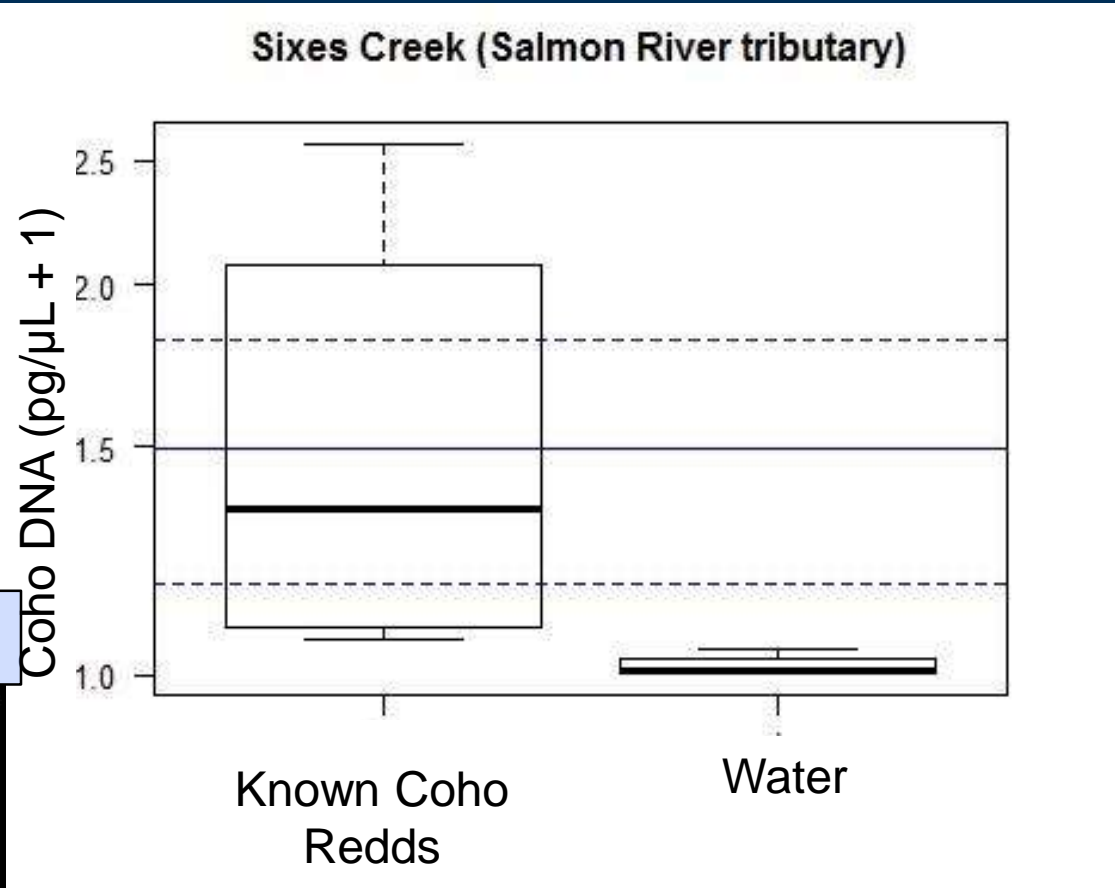
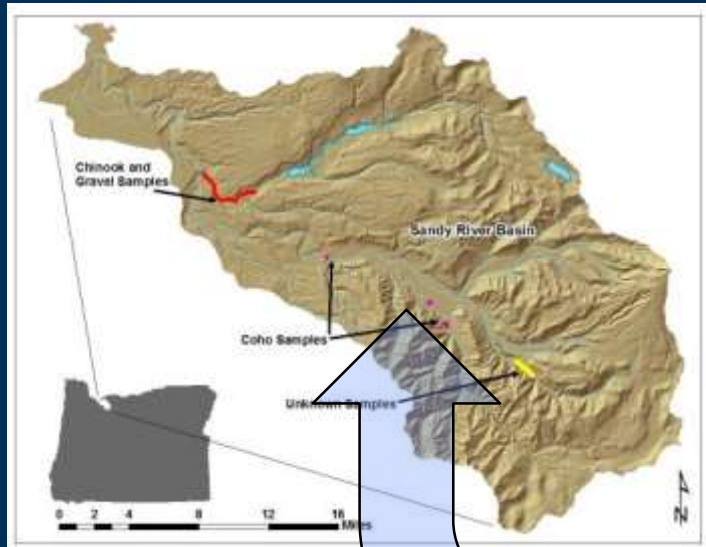




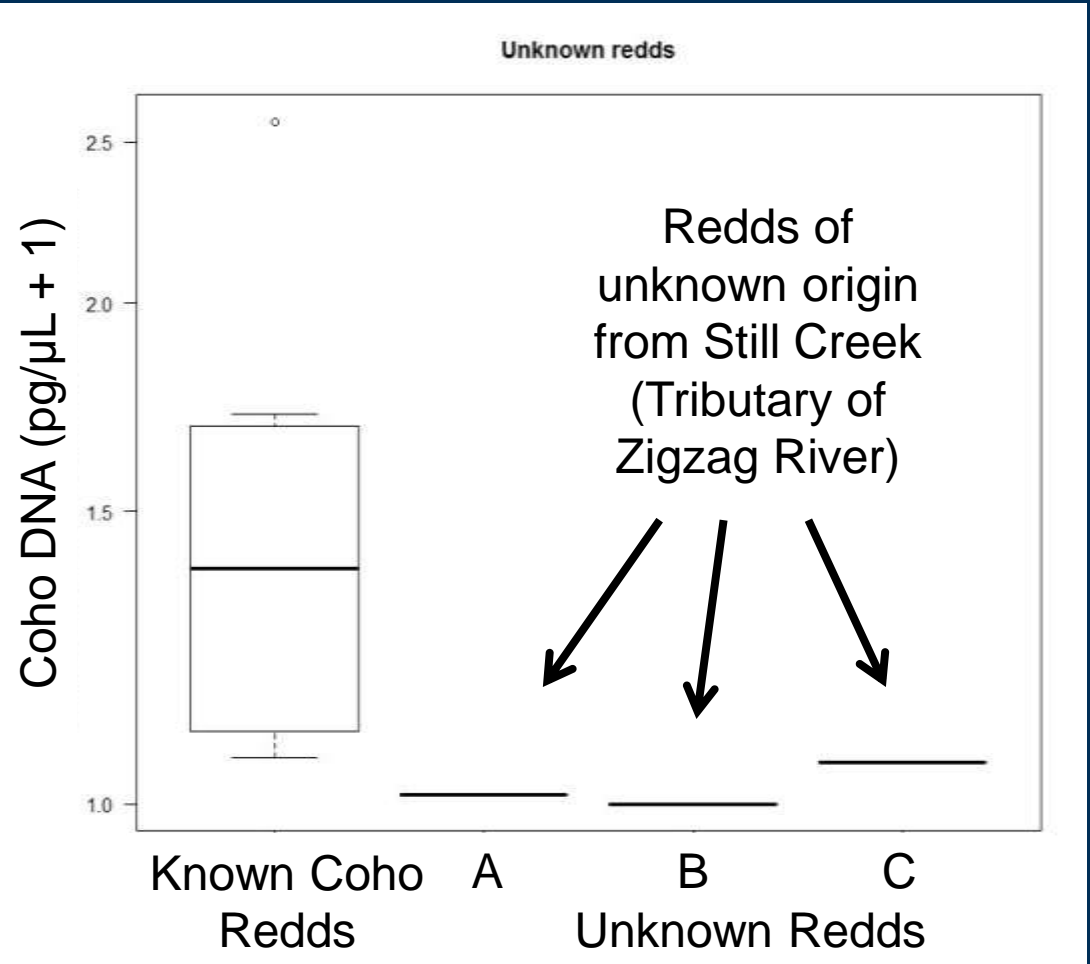
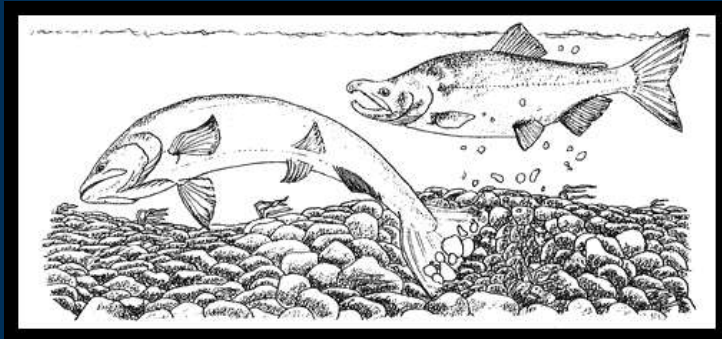
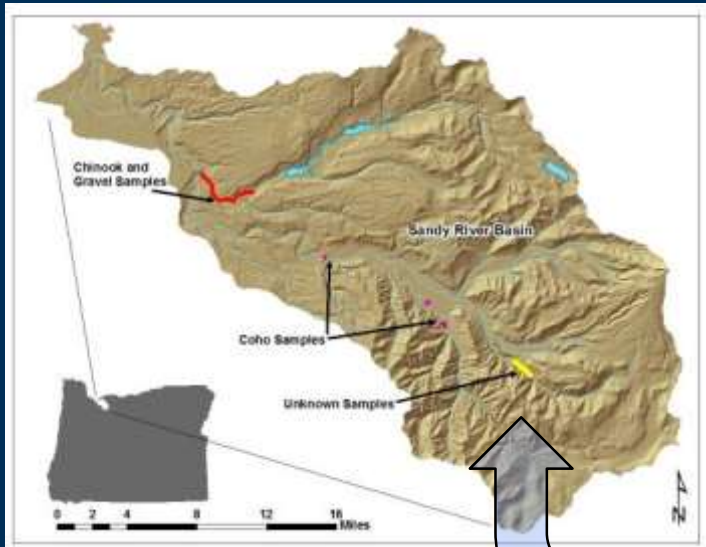
1. 15 mL water samples (triplicate)
2. Field preserved with 1.5 mL sodium acetate and 33 mL ethanol
3. DNA extracted via precipitation method (Ficetola et al. 2008)
4. qPCR analysis



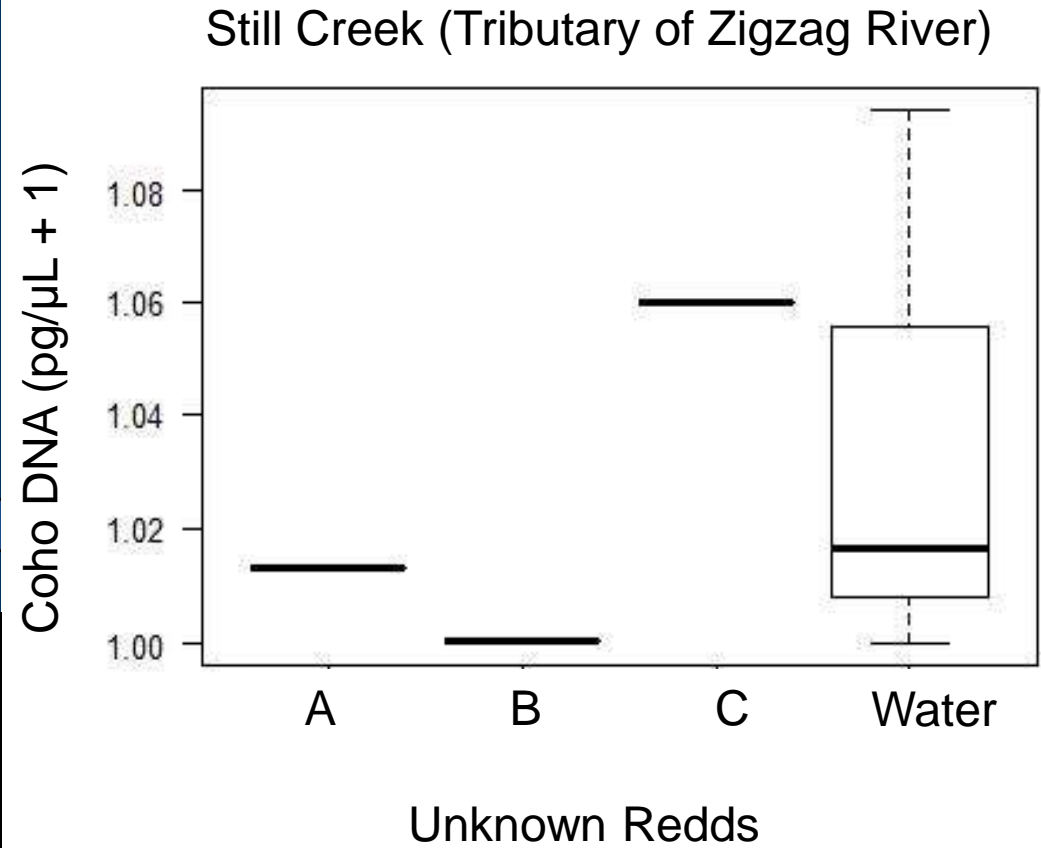
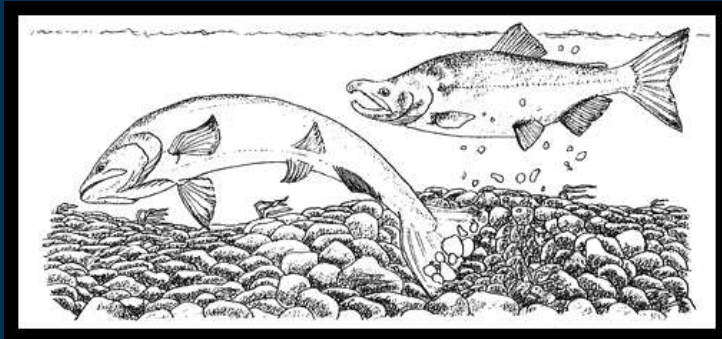
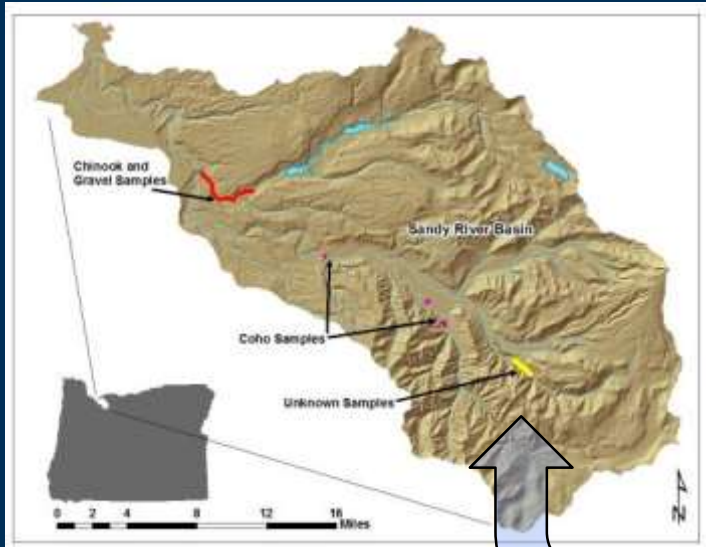
# How much Coho DNA is at a Coho Redd?



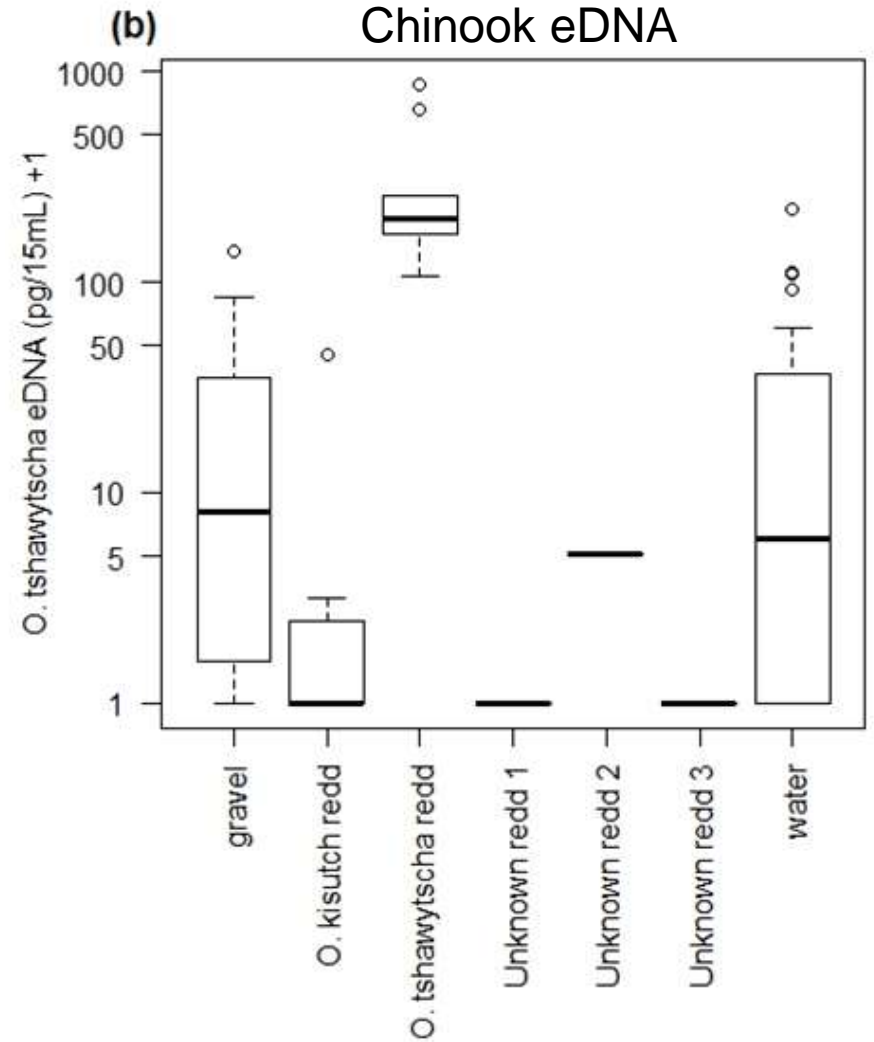
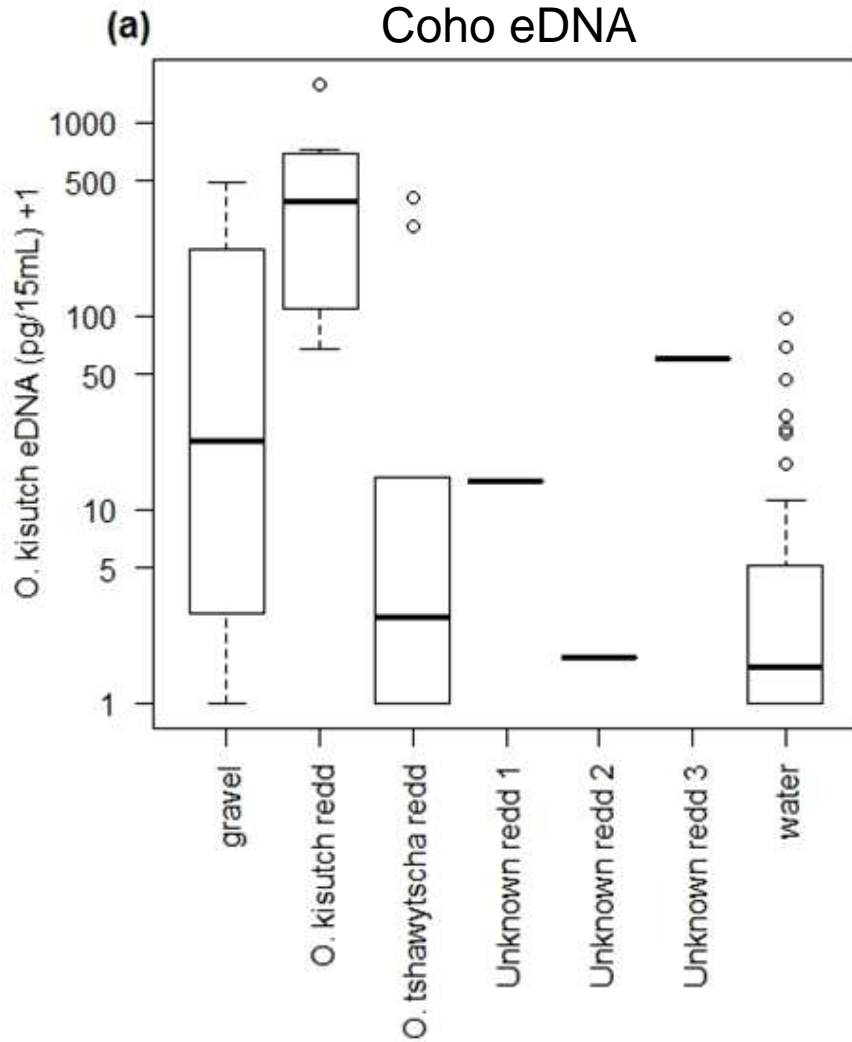
# Can unknown redds be assigned?



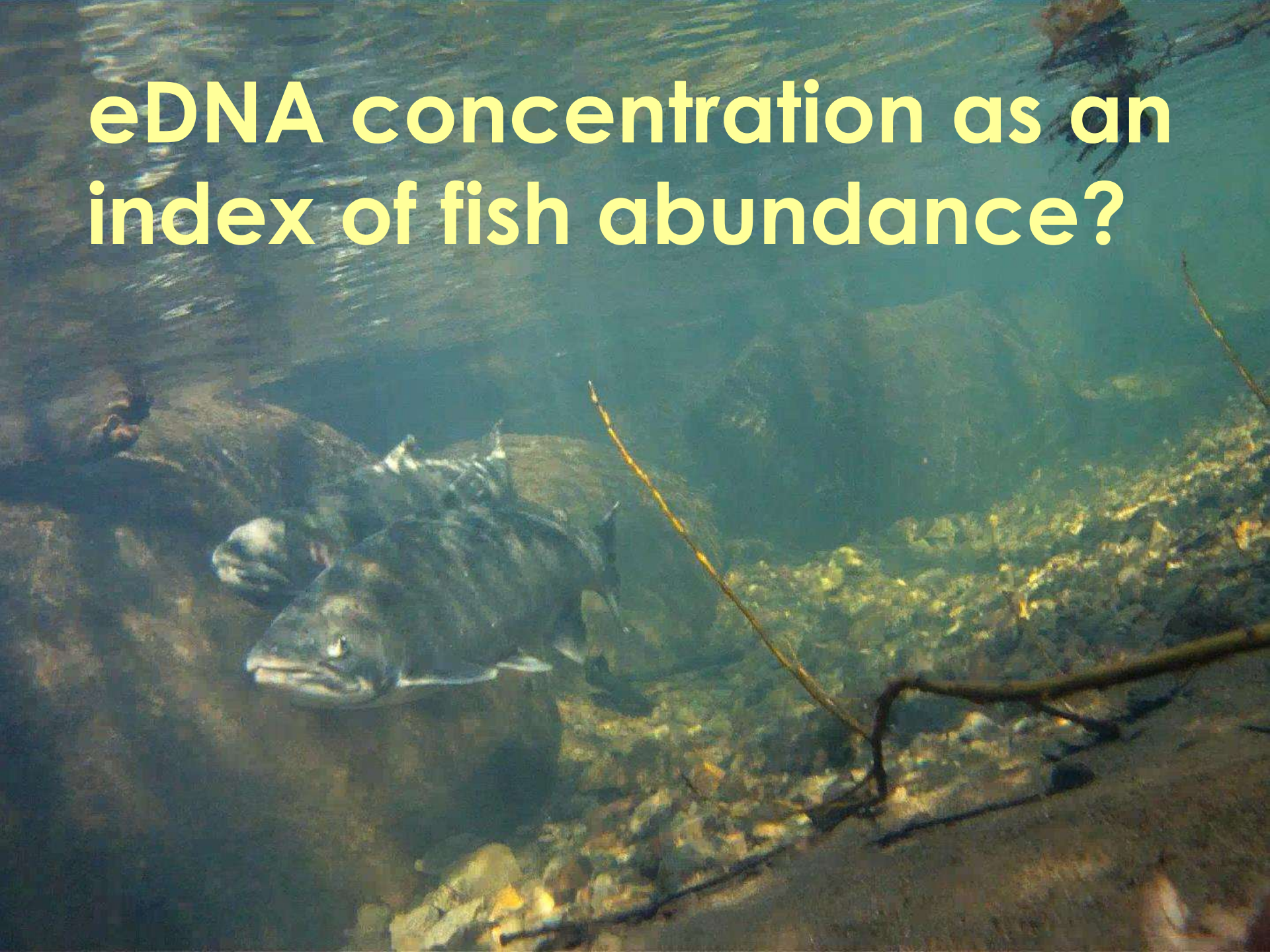
# Can unknown redds be assigned?



# All streams combined



**eDNA concentration as an  
index of fish abundance?**



# Omak Creek

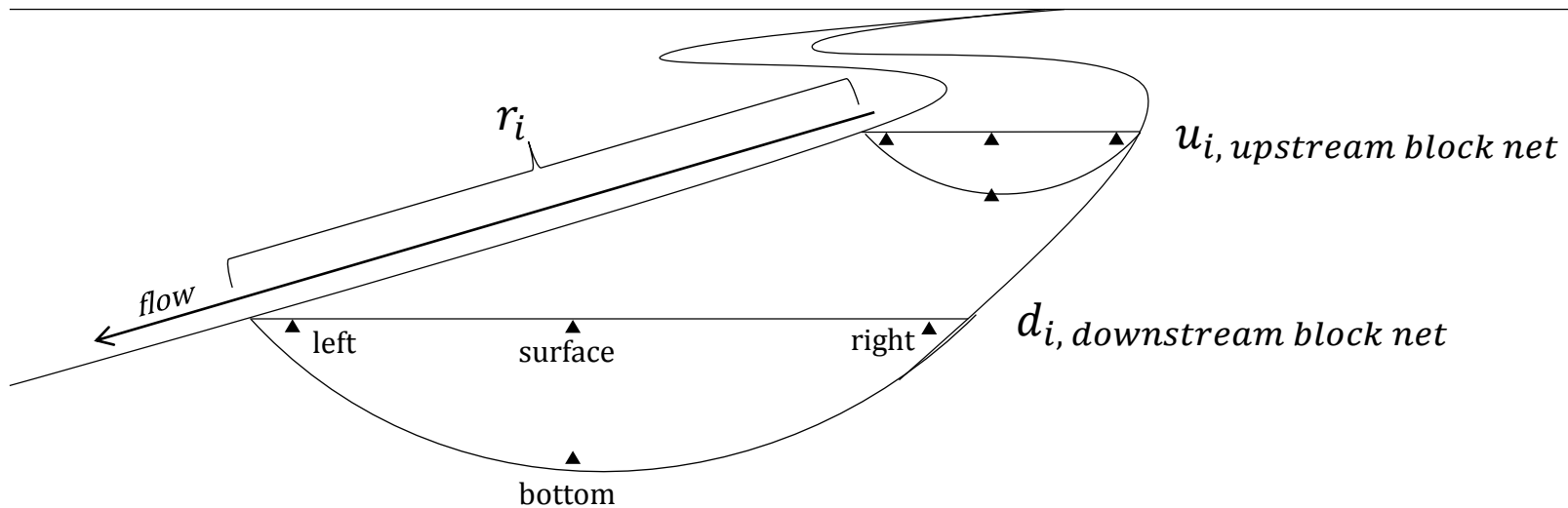


- Mid-size perennial stream
- ~5 m wetted width
- 10 - 150cfs
- USGS Gage  
12445900

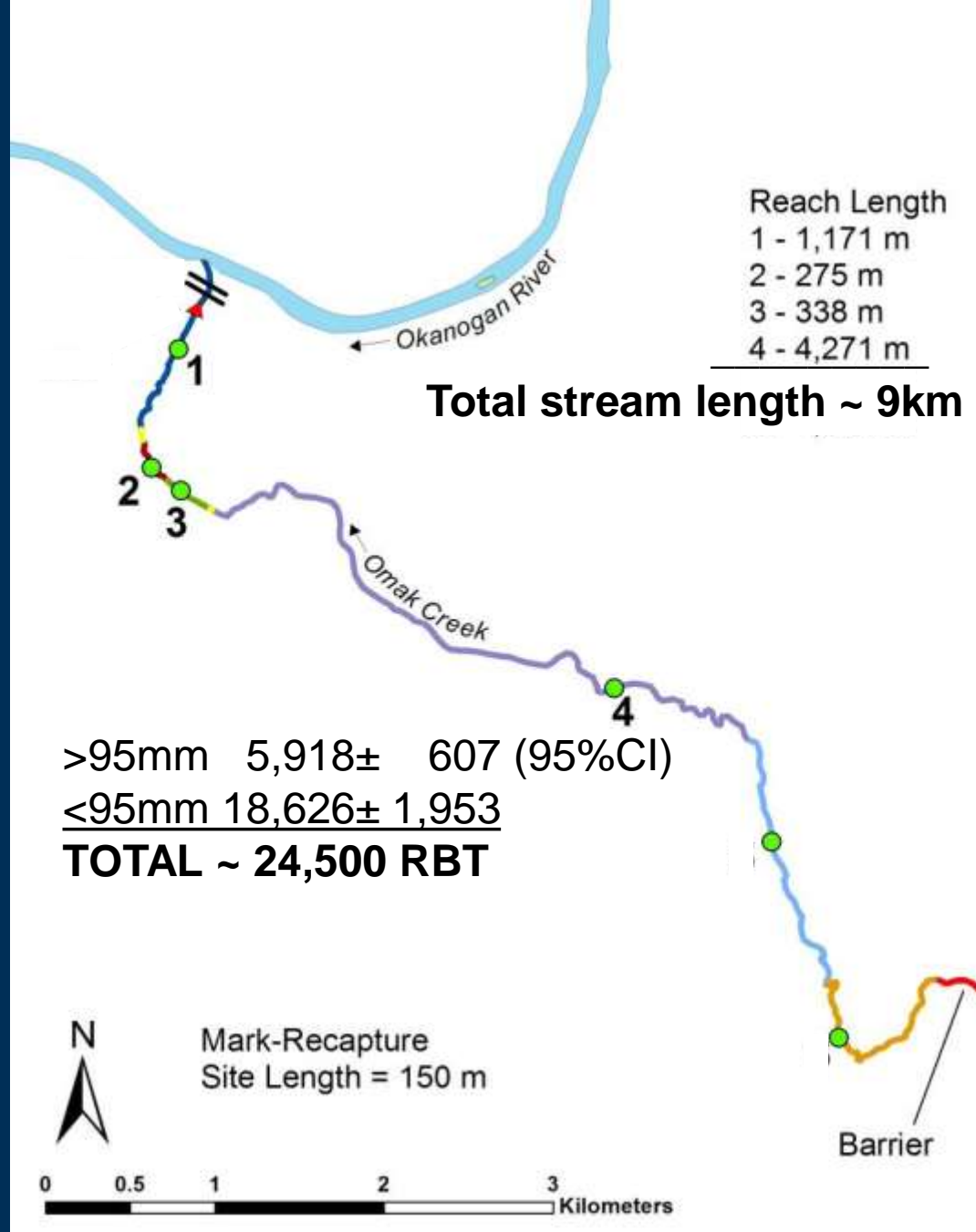


1. Does [eDNA] reflect relative fish abundance?
2. Does it matter where samples are collected (cross-section)?

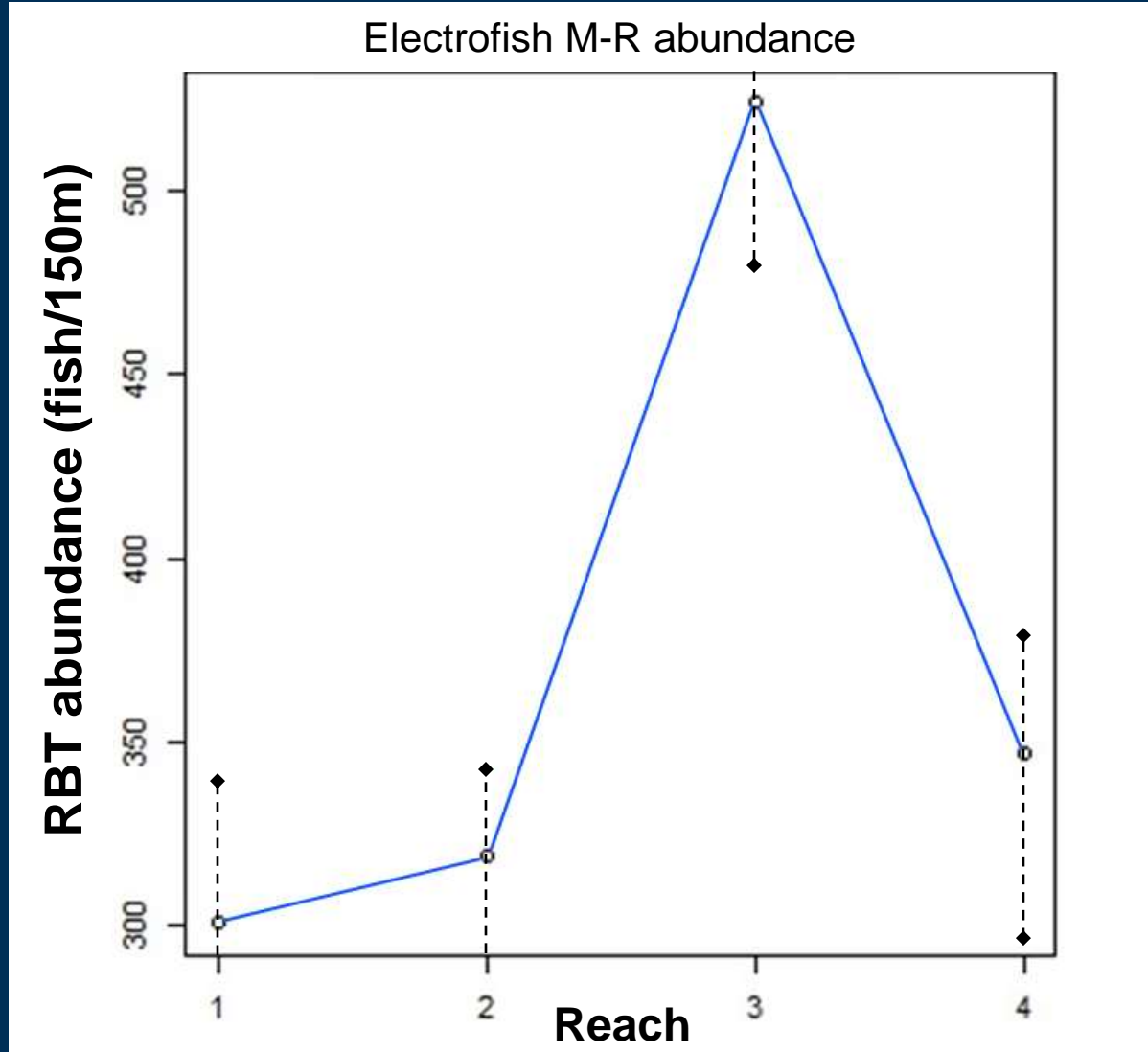
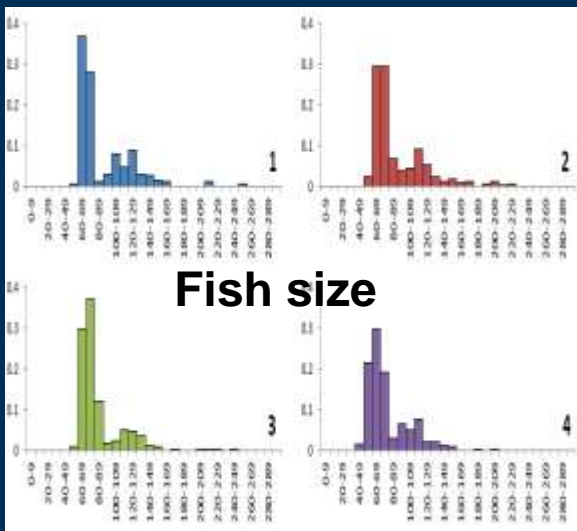
*Reach length eDNA (diff.);  $r_i = \bar{d}_i - \bar{u}_i$*



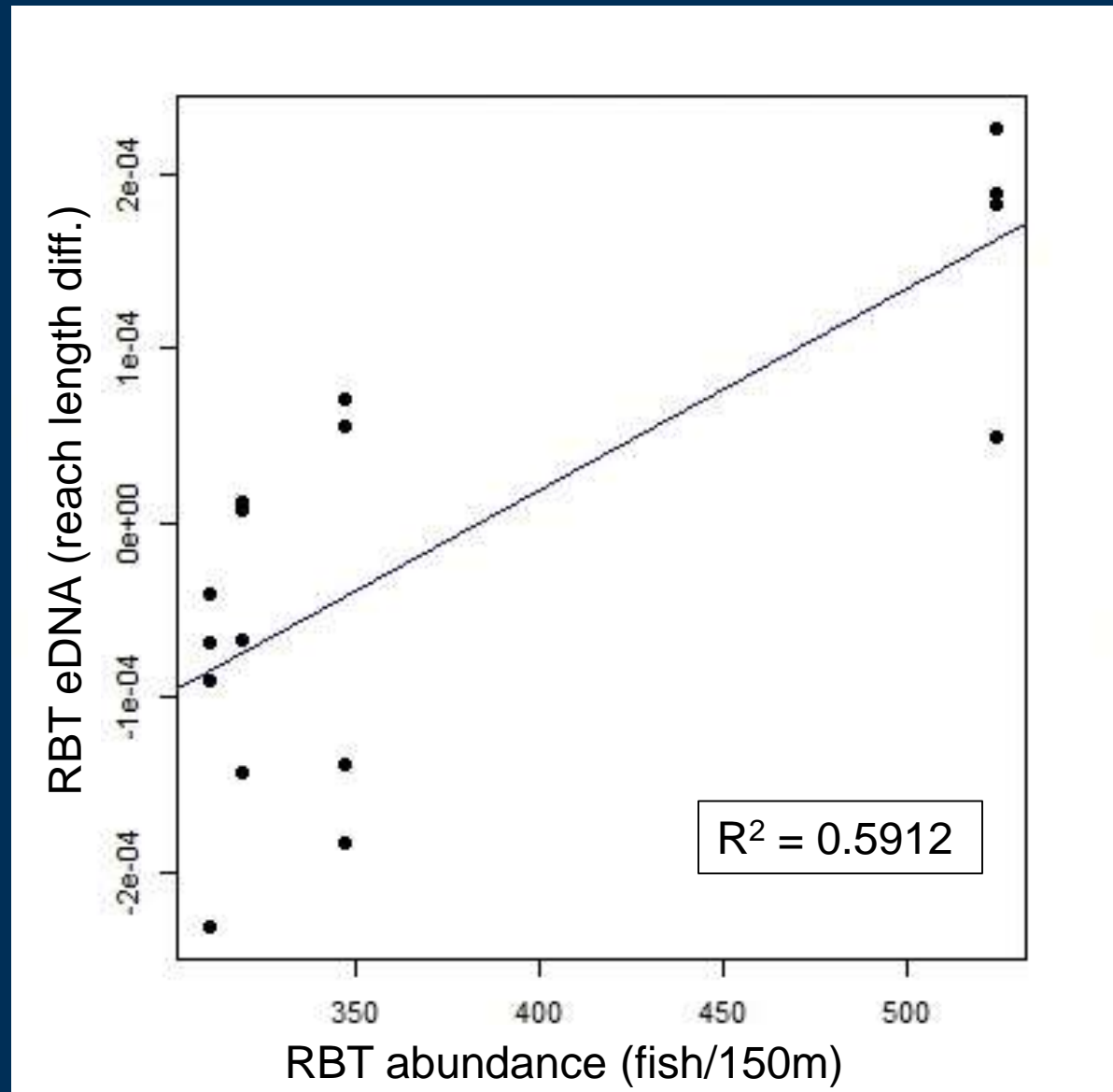




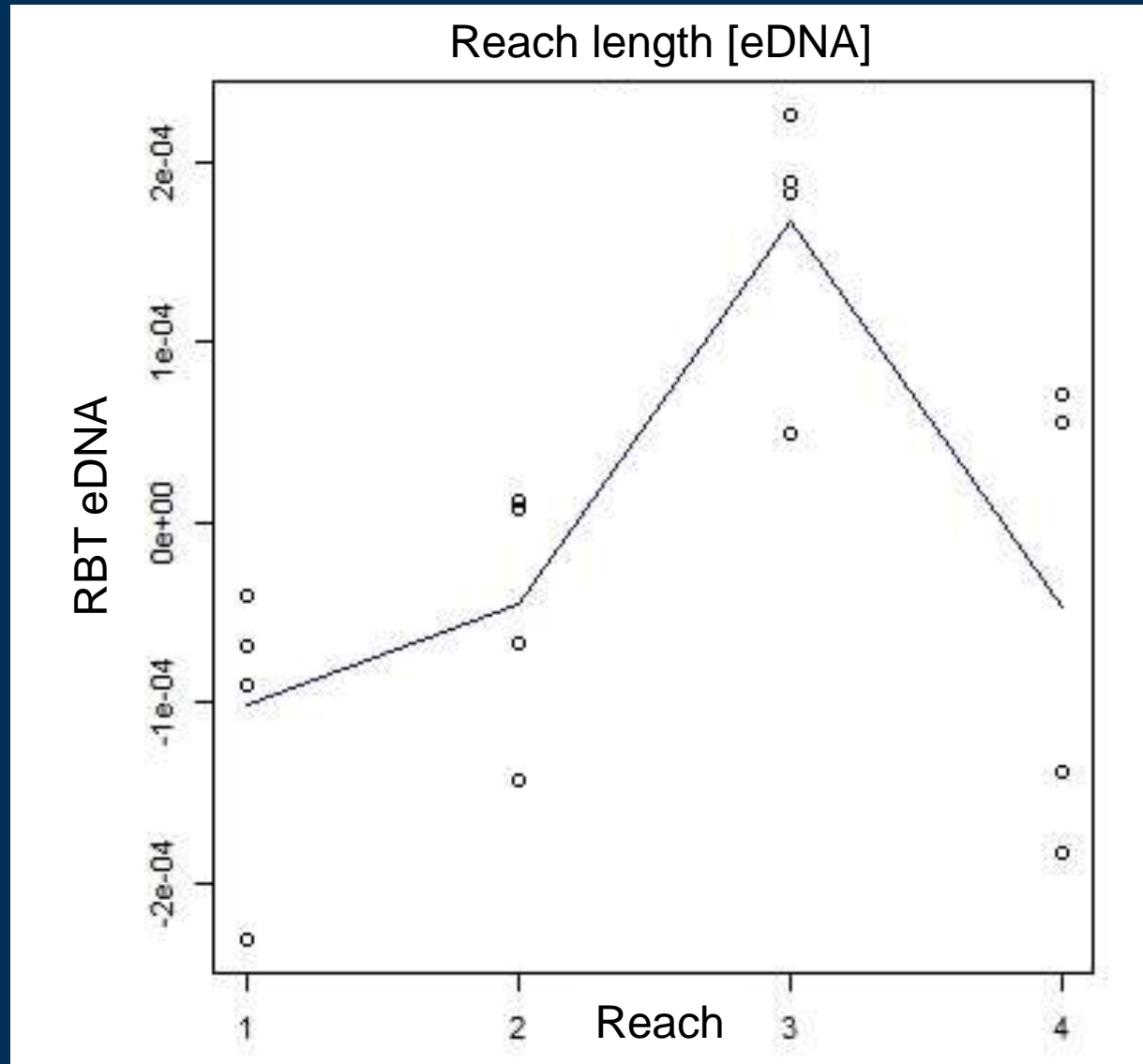
# Electrofishing mark-recap RBT abundance



# Does [eDNA] reflect fish abundance?

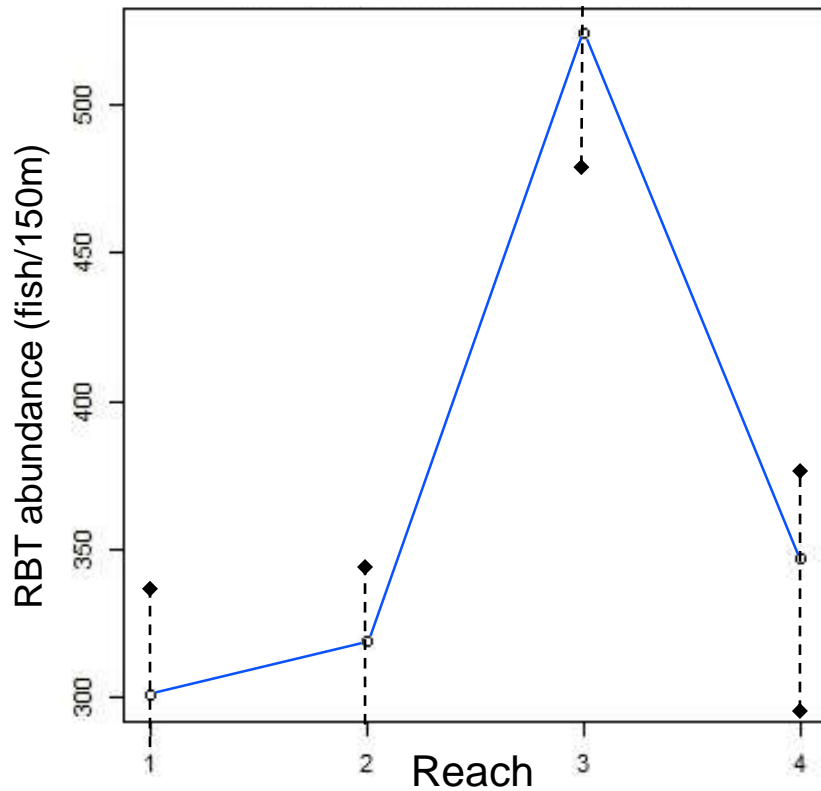


# eDNA as an index of relative abundance

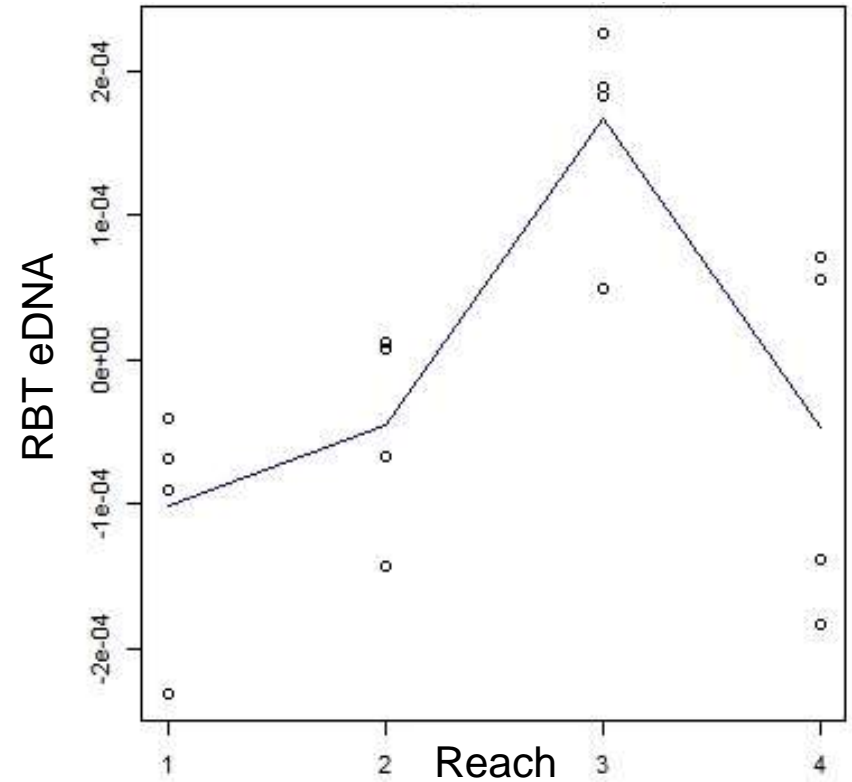


# eDNA as an index of relative abundance

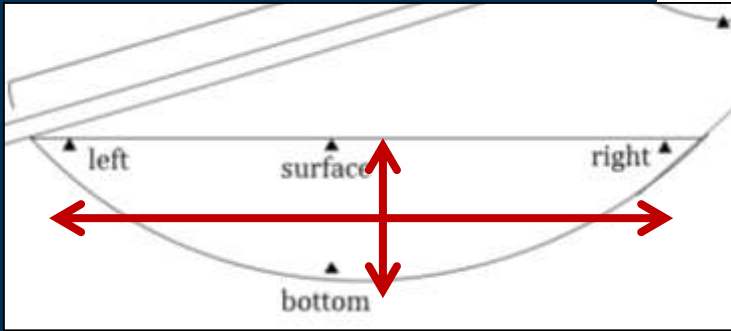
Electrofishing M-R abundance



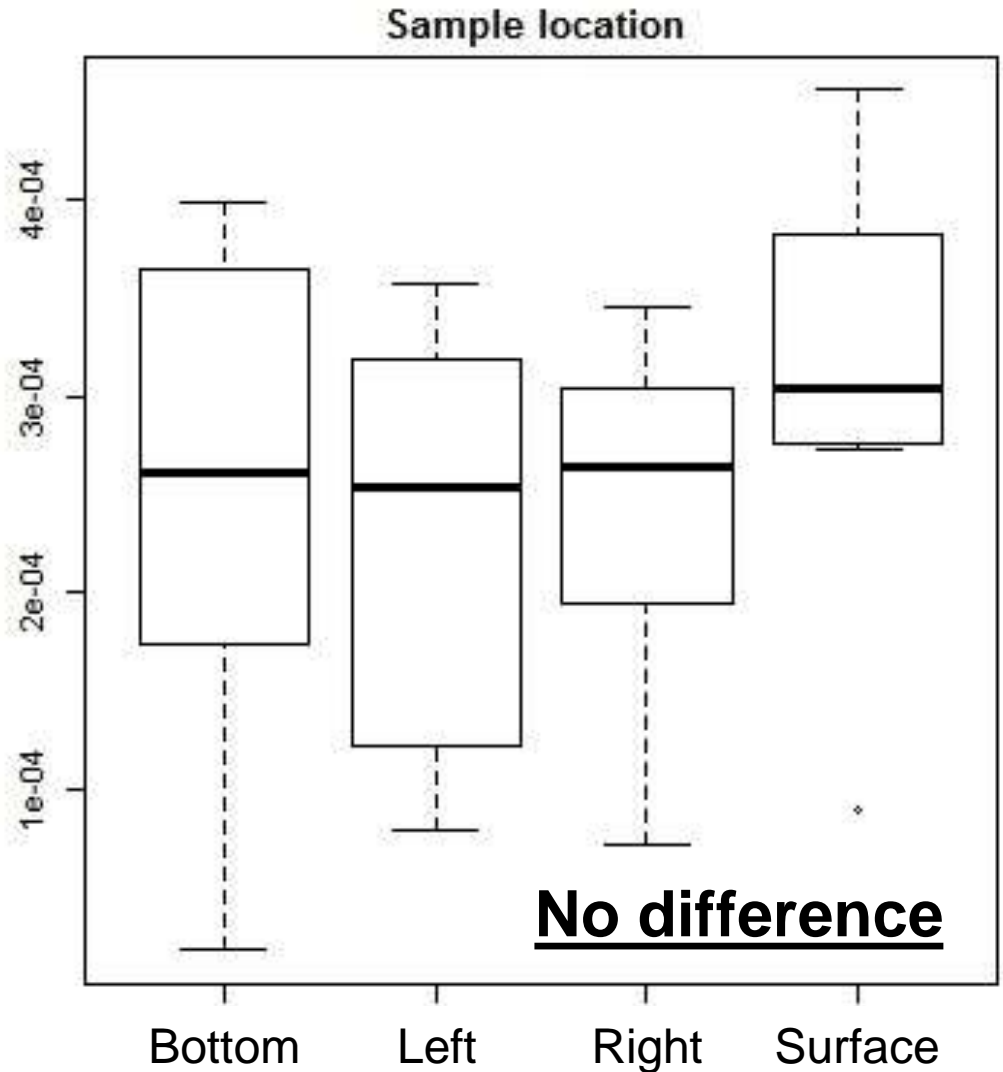
Reach length [eDNA]



# eDNA within the stream cross-section



RBT eDNA



# eDNA Sampling Protocols



Prepared in cooperation with Washington State University

## Environmental DNA Sampling Protocol—Filtering Water to Capture DNA from Aquatic Organisms

Chapter 13 of  
Section A, Biological Science  
Book 2, Collection of Environmental Data

Techniques and Methods 2–A13

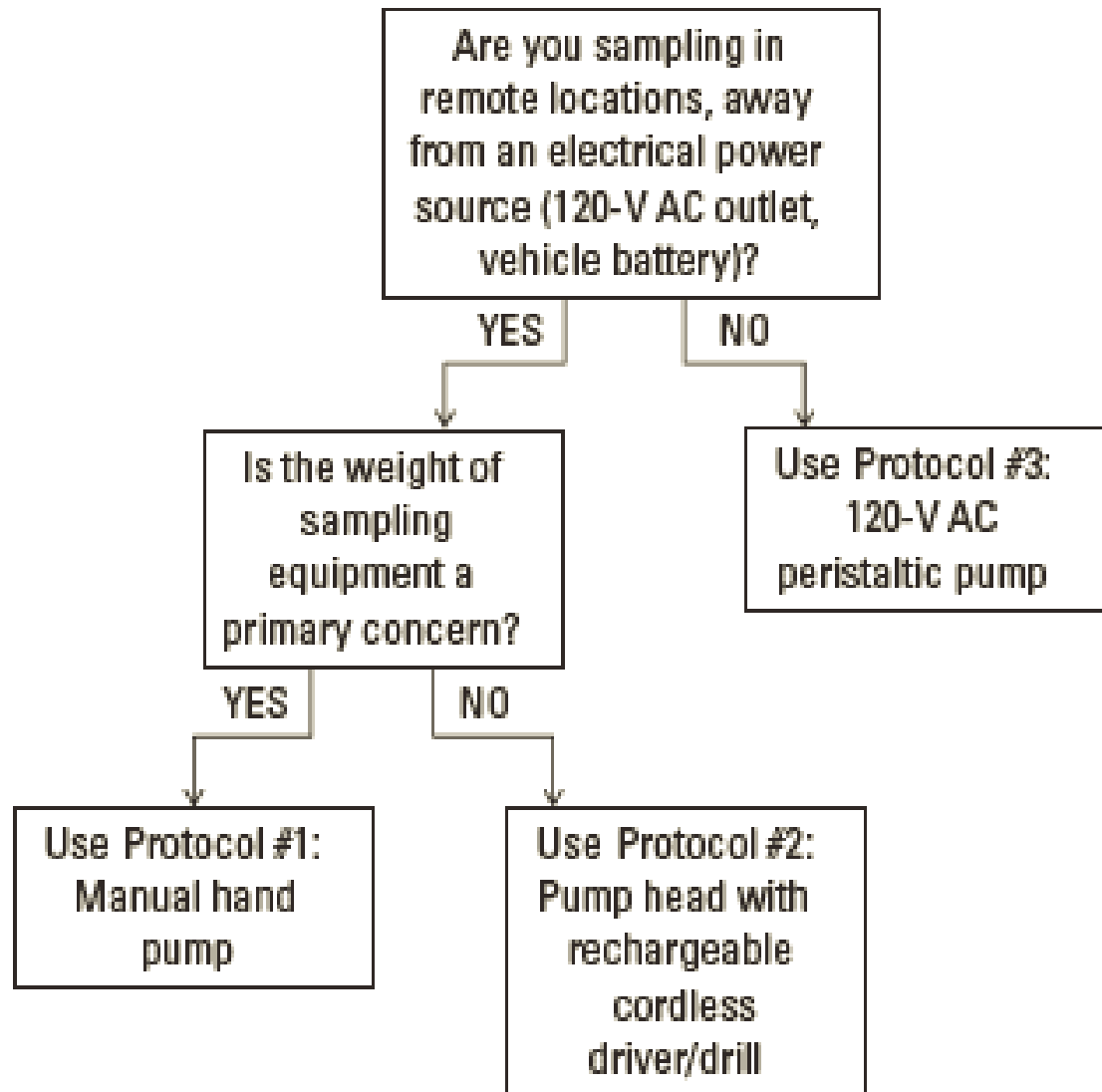
U.S. Department of the Interior  
U.S. Geological Survey



<http://pubs.usgs.gov/tm/02/a13/tm2a13.pdf>



# Selecting the best protocol





# Sampling Workflow Diagram

Step 1: Choose the best protocol, depending on your conditions



Protocol #1: Hand pump



Protocol #2: Cordless driver



Protocol #3: 120-V pump

Step 2: Collect water sample



Direct

or



Collect and pour

Step 3: Preserve water sample



Ethanol-filled vial



# Protocol #1: Hand pump



# Protocol #2: Cordless driver

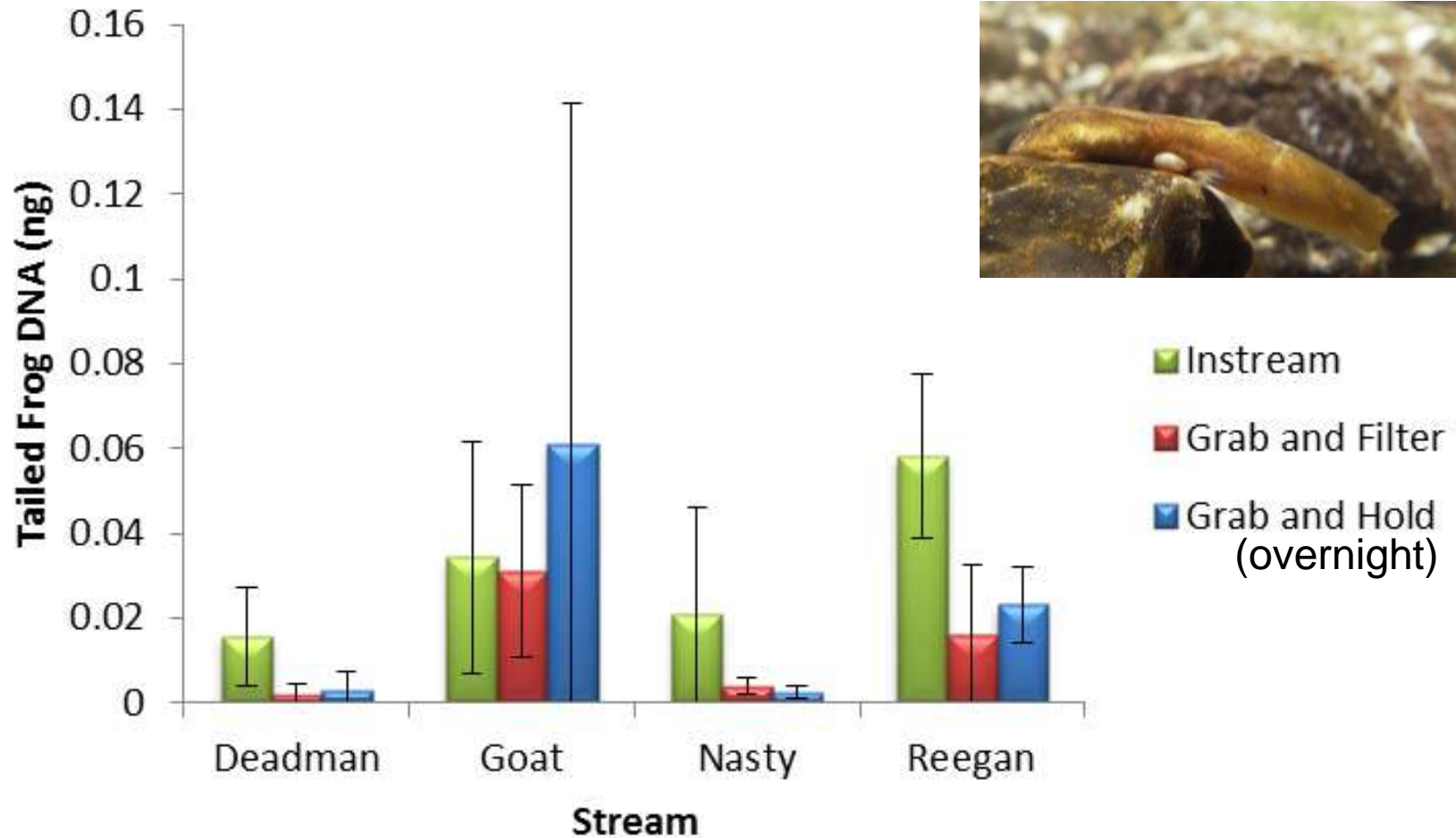


# Protocol #3: 120v pump

# Sample collection options

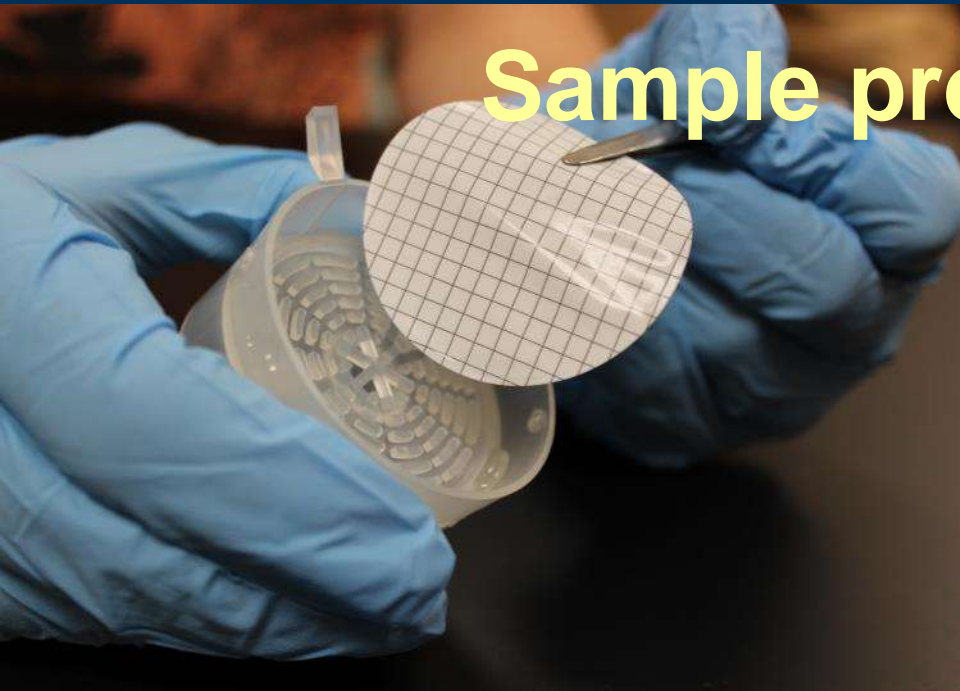


# Comparing sample collection options



$P = 0.02$  Instream samples higher

# Sample preservation



Filters  
stored in  
ethanol at  
room temp

# eDNA Resources



Prepared in cooperation with Washington State University

## Environmental DNA Sampling Protocol—Filtering Water to Capture DNA from Aquatic Organisms

Chapter 13 of  
Section A, Biological Science  
Book 2, Collection of Environmental Data

## SAMPLING PROTOCOLS

<http://pubs.usgs.gov/tm/02/a13/tm2a13.pdf>



## Application of Environmental DNA for Inventory and Monitoring of Aquatic Species

*This fact sheet was created to help biologists and resource managers understand emerging methods for detecting environmental DNA and their potential application for inventorying and monitoring aquatic species. It is a synthesis of published information.*



## USGS FACT SHEET

<http://pubs.usgs.gov/fs/2012/3146/pdf/fs2012-3146.pdf>





# eDNA Resources



[eDNA.fisheries.org](http://eDNA.fisheries.org)

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Photo courtesy of Jeffrey Williams (AKDFG)

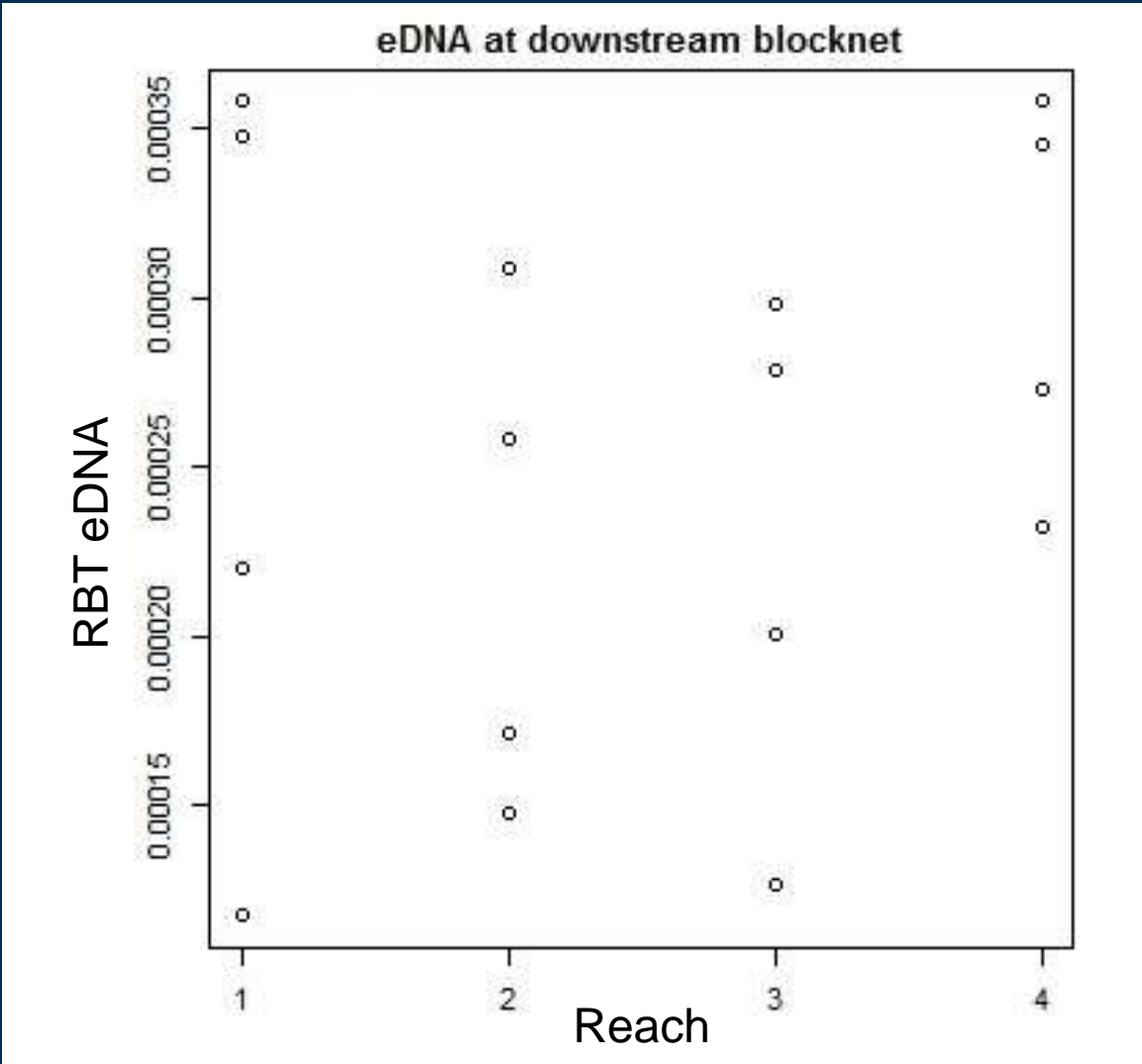


# Acknowledgements

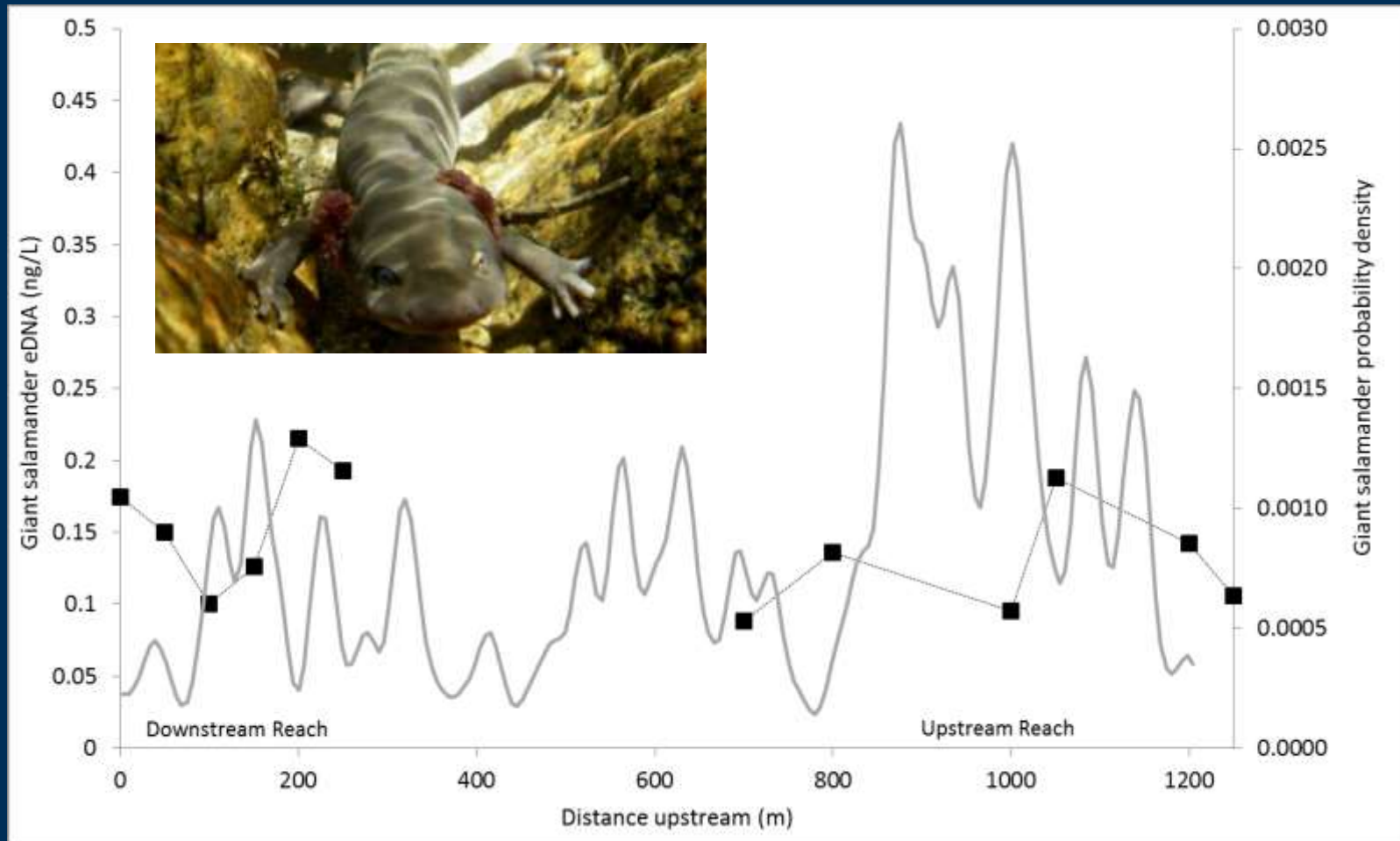
- **Portland Water Bureau (PWB)**  
**Burke Strobel**
- **Colville Confederated Tribes (CCT)**  
**Okanogan Basin Monitoring & Evaluation Program (OBMEP)**  
**Chief Joseph Hatchery Science Program (CJHP)**
- **Washington State University (WSU)**  
**Caren Goldberg & Kath Strickler**





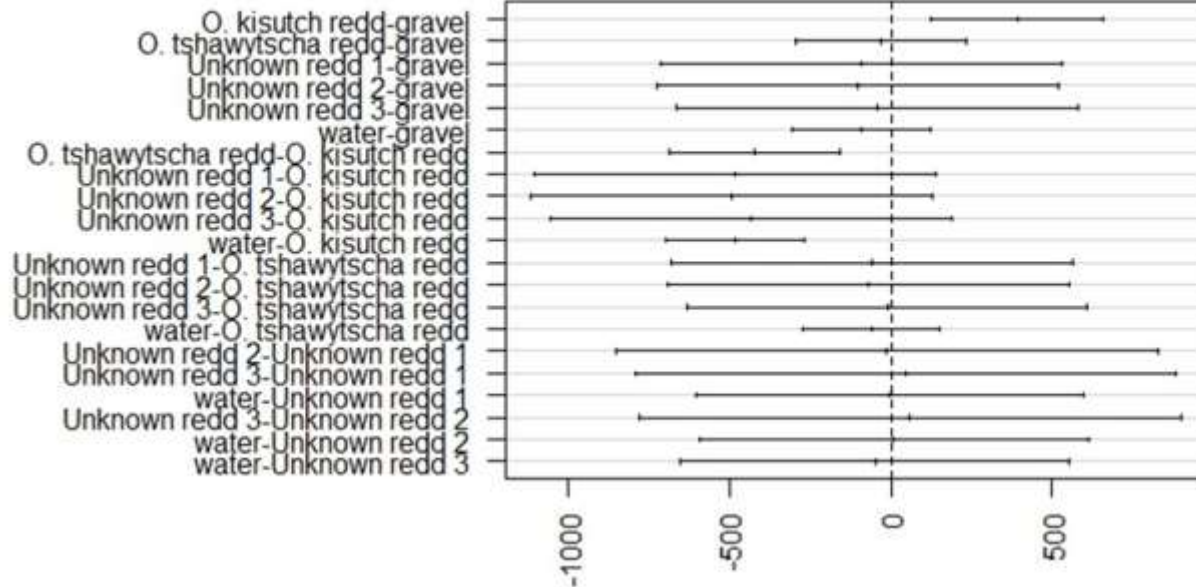


# What does an eDNA sample represent?



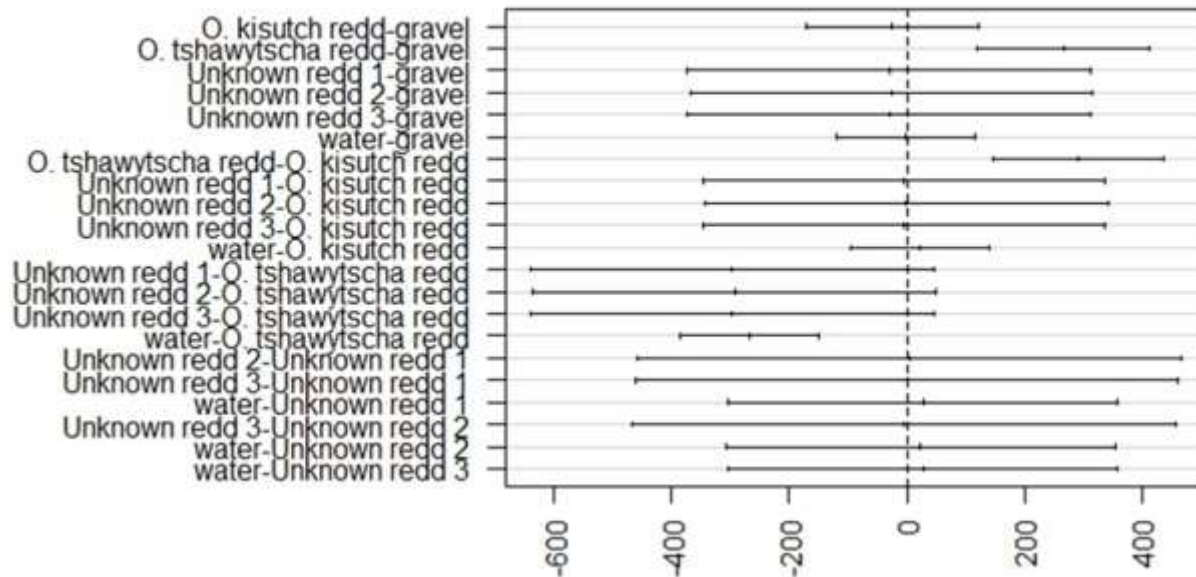
### (a) Coho eDNA

95% family-wise confidence level



### (b) Chinook eDNA

95% family-wise confidence level



**Table 3. Analysis of Variance (ANOVA) table for differences in *O. kisutch* eDNA among sample types.**

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
Sample type	6	1847497	307916	8.1409	1.997e-06 ***	
Residuals	59	2231569	37823			
Signif. codes:	0 '****'	0.001 '**'	0.01 '*'	0.05 '.'	0.1 ' '	1
Response: Site replicate mean [eDNA] (pg/15 mL)						

**Table 4. Tukey multiple comparisons of means w/ 95% family-wise confidence level for *O. tshawytscha* eDNA among sample types.**

Sample type	diff	lwr	upr	p adj
<i>O. tshawytscha</i> redd - gravel	266.7449	120.5793	412.91037	0.0000133
<i>O. tshawytscha</i> redd - <i>O. kisutch</i> redd	291.8099	145.6444	437.97538	0.0000018
Water - <i>O. tshawytscha</i> redd	-269.2079	-387.1876	-151.22823	0.0000001
Fit: aov(formula = Site replicate mean [eDNA] (pg/15mL)~ Sample type, data = <i>O. tshawytscha</i> eDNA)				



# How many replicates are necessary?

Possible detection outcomes at a site	% of sites	# replicates (-)	# replicates total
0 0 0	41%		
1 0 0	3%	6	9
1 1 0	7%	6	18
1 1 1	49%	0	135
Total	100%	12	162

# Assessing detection probability and error

Possible detection outcomes at a site	% of sites	# replicates (-)	# replicates total
0 0 0	41%		
1 0 0	3%	6	9
1 1 0	7%	6	18
1 1 1	49%	0	135
Total	100%	12	162

$12/162 = 7\%$  false negatives



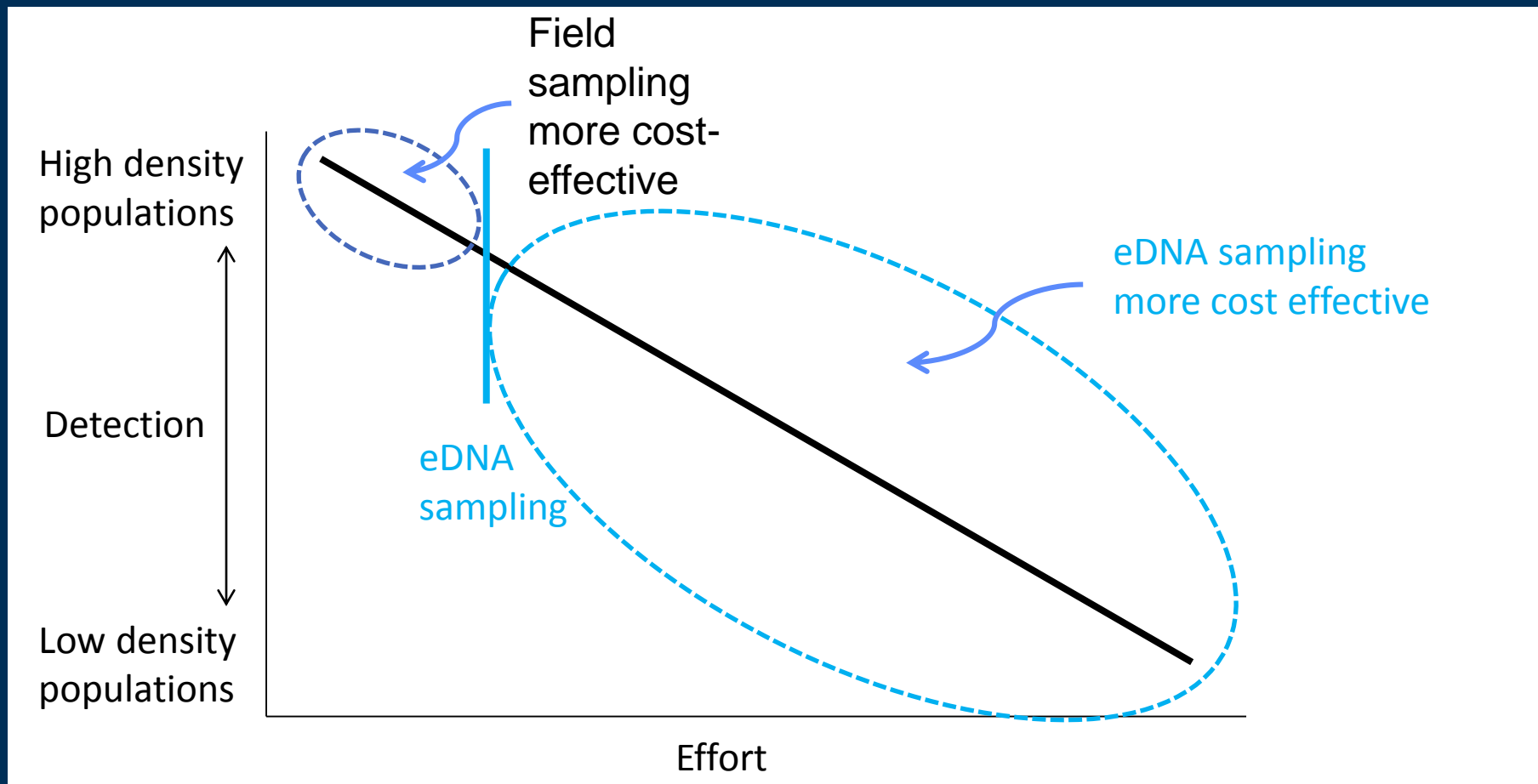
## Contamination Prevention

Contamination can result from various factors at every step in the sample collection process. Be vigilant. Before initiating eDNA sample collection, the following field and laboratory practices should be reviewed to avoid contamination of samples and cross-contamination among samples:

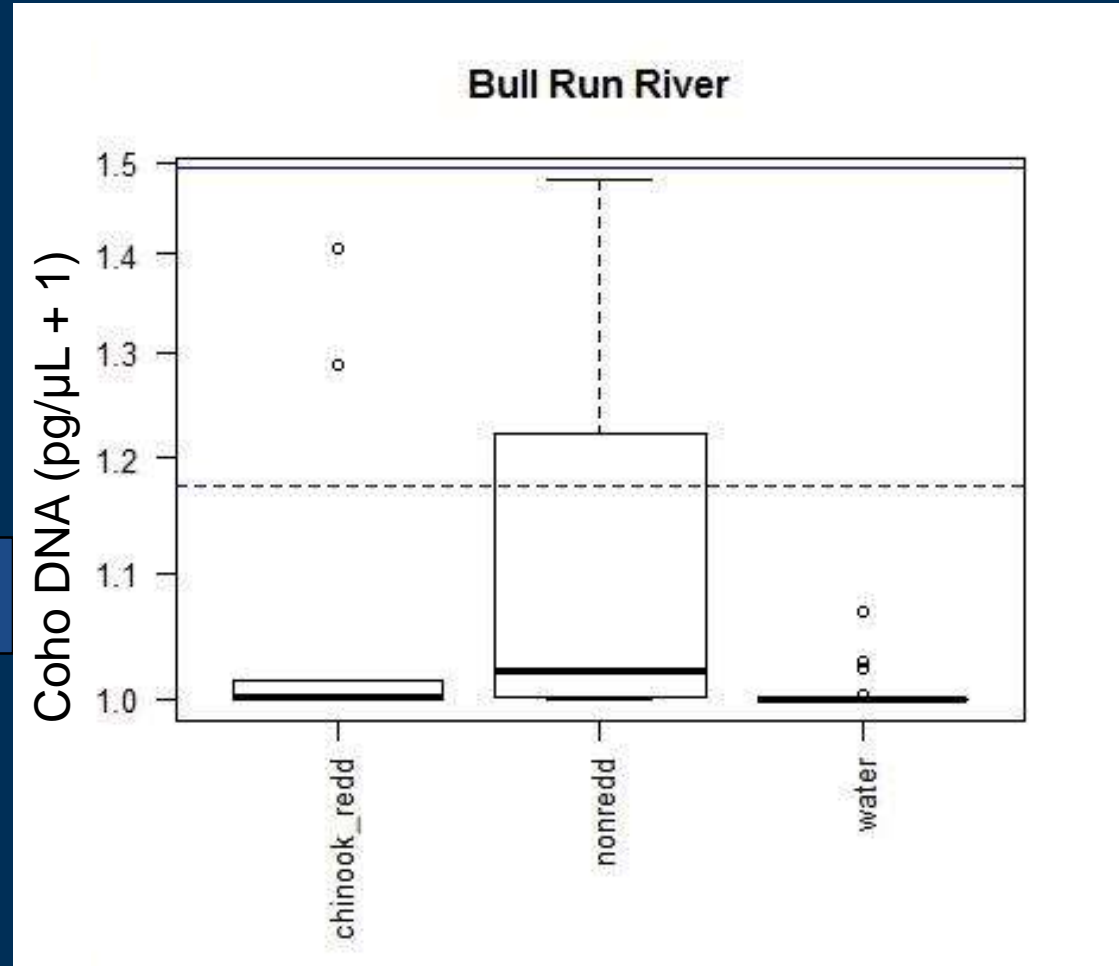
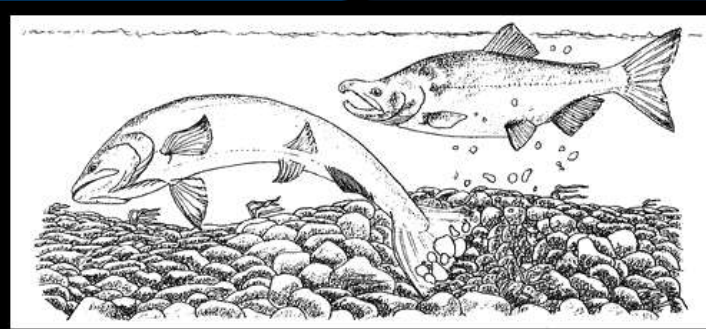
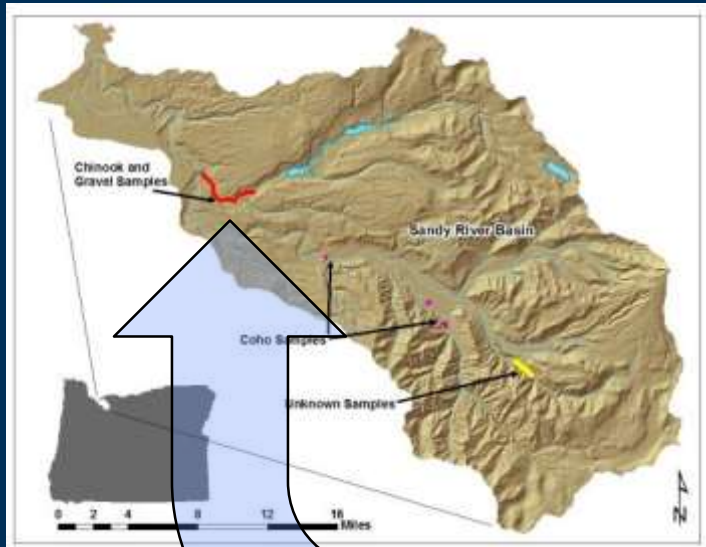
- Wear clean, non-powdered, single-use gloves when collecting samples and removing filters. Do not let gloves contact contaminated surfaces, such as any equipment that was not sterilized between sites, prior to handling the filter.



# When do we use eDNA?



# How much Coho DNA is in environment?



# How much Coho DNA is at a Coho Redd?

